

Three new species and one new record of Deimatidae (Echinodermata, Holothuroidea, Synallactida) discovered in the South China Sea and the Mariana fore-arc area using integrative taxonomic methods

Yunlu Xiao^{1,2}, Haibin Zhang¹ 

¹ Institute of Deep-sea Science and Engineering, Chinese Academy of Sciences, Sanya 572000, China

² University of Chinese Academy of Sciences, Beijing 100049, China

Corresponding author: Haibin Zhang (hzhang@idsse.ac.cn)



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Abstract

Deep-sea holothurian specimens were collected during five scientific expeditions (2018–2023) using the submersible vehicle ‘Shenhaiyongshi’. Our examination of specimens of Deimatidae from the South China Sea and the Mariana fore-arc area revealed three new species, which were described as *Oneirophanta idsseica* **sp. nov.**, *Oneirophanta brunneannulata* **sp. nov.**, and *Oneirophanta lucerna* **sp. nov.** These species were distinguished from each other and from congeners by the arrangement, and number of ventrolateral tube feet and ossicle types. We also reported *Oneirophanta mutabilis mutabilis* Théel, 1879 for the first time from the Mariana fore-arc area, and we recorded *Deima validum validum* for the second time from the South China Sea. The taxonomy of these new species and new records is discussed, and a phylogenetic analysis based on a concatenated dataset of 16S and COI genes was conducted. Additionally, the inter- and intraspecific genetic divergences we calculated among deimatid species. The results support the assignment of these new species to the genus *Oneirophanta* and their separation from congeners. A description of the main morphological characters of *Oneirophanta* species is also provided. The data were collected from geographically diverse areas and suggest that species of Deimatidae were abundant in the Pacific Ocean and occupied a wide range of depths.

Key words: COI, deep-sea, morphology, *Oneirophanta*, phylogeny, sea cucumber, SEM, taxonomy

Introduction

Echinoderms are abundant in Chinese seas, and the South China Sea has more species than the Yellow Sea and the East China Sea (Liao and Xiao 2012). The South China Sea covers ~ 3.5 million km², and it is studded with 200 islands and islets. It is in the center of the Indo-Western Pacific Biogeographic Province, and it is one of the most biodiverse seas in the world. The maximum recorded depth for echinoderms was 5377 m, with considerable habitat and species diversity (Morton and Blackmore 2001; Teh et al. 2019). Deep-sea exploration in

China began with the *Jiaolong* manned submersible in 2010, which has been used widely to uncover the deep-sea biodiversity in China seas and the Mariana Trench (Li 2017). Taxonomic research has revealed many new species and new geographical records of animal species in the deep waters of the South China Sea (Li et al. 2019).

Holothurians are the dominant epibenthic invertebrate taxon in many areas of the deep sea, and they account for 90% of that ecosystem's biomass (Hendler et al. 1995). However, our present knowledge of deep-sea holothurians is still limited due to the difficulty in collecting and depositing good specimens. Even the specimens collected by the submersible carefully and carried from the seabed to the sea surface in seawater-filled containers have arrived aboard the ship in very poor condition (Pawson 1982a). The epidermis and dermis of the whole holothurians begin to slough off, and the entire external body wall is often completely autotomized. The body wall contains a high proportion of water, even the most carefully preserved specimens will shrink greatly, and the shrinkage rate is frequently greater than 90% (Billett 1991). When preserved in absolute ethanol, the holothurians shrink, so all the external characters are easily distorted, which undoubtedly impacts their morphological identification negatively after collection. In recent years, few studies on the order Elasipodida have been conducted in the South China Sea (Li et al. 2018; Xiao et al. 2018, 2023), and studies on other taxa in the South China Sea have also been reported rarely. More morphological information and molecular data should be obtained for a comprehensive taxonomic evaluation of deep-sea holothurians in the South China Sea (Li et al. 2019).

The family Deimatidae formerly belonged in the order Elasipodida Théel, 1882, but was later transferred to the order Aspidochirotida by Smirnov (2012). Miller et al. (2017) subdivided paraphyletic ordo Aspidochirotida into three separate orders and placed family Deimatidae to the order Synallactida Miller, Kerr, Paulay, Reich, Wilson, Carvajal & Rouse, 2017. The order Synallactida includes the families Deimatidae Théel, 1882, Stichopodidae Haeckel, 1896, and Synallactidae Ludwig, 1894. Deimatidae is the smallest family in the order, and it contains 13 accepted species within three genera (*Oneirophanta* Théel, 1879, *Orphnurgus* Théel, 1879, and *Deima* Théel, 1879) (WoRMS 2023). Only two of the recognized species in the family Deimatidae, *Orphnurgus protectus* (Sluiter 1901b) and *Deima validum* Théel, 1879, have been recorded from the South China Sea (Liao 1997). The genus *Oneirophanta* was established with *Oneirophanta mutabilis* Théel, 1879 as the type species (Théel 1879). Currently, there are only three species of *Oneirophanta*, and none of them have been found in China. Eleven deimatid specimens were collected from the South China Sea and the Mariana fore-arc area from 2018 to 2023. Morphological observations of these specimens suggested that they represented three new species and one new record from the Mariana fore-arc area, and they all belonged to the genus *Oneirophanta*.

In addition, we present a morphological description of *Deima validum validum* Théel, 1879, which was recorded for the second time in the South China Sea; the present specimens show some variations compared with specimens that were recorded previously. Our study provides comprehensive a description of morphological characters, an assessment of intraspecific divergence between the new species and all other known species, and more molecular details that may be useful for further studies of the phylogeny and diversity of the family Deimatidae.

Materials and methods

Sampling and preservation

Specimens were collected from the South China Sea and the Mariana fore-arc area (Fig. 1) using the manned submersible vehicle ‘Shenhaiyongshi’ from 2018 to 2023, at depths of 1340–3806 m. Samples were frozen or preserved in absolute ethanol, and then stored at the Institute of Deep-sea Sciences and Engineering (IDSSE), Chinese Academy of Sciences (CAS), Sanya, China.

Morphological observations

The specimens of each species were identified using a variety of original descriptions and literature (Théel 1879; Koehler and Vaney 1905; Hansen 1967, 1975; Pawson 2002). External morphological features were examined under a dissecting stereomicroscope (OLYMPUS SZX7), and identification was based on in situ images or pictures photographed in our lab using a Canon EOS 6DII camera. To prepare the deposits, small pieces of body tissue (dorsal and ventral body wall, tentacles, papillae, and tube feet) were digested in a 15% sodium hypochlorite solution. The deposits were then washed three times in distilled water and dried in absolute ethanol before examination with a scanning electron microscope (Phenom ProX).

DNA extraction, PCR amplification, and DNA sequencing

Total genomic DNA was extracted from small pieces of 20–30 mg holothuri-an muscle tissue using a TIANamp Marine Animals DNA Kit (TianGen, Beijing),

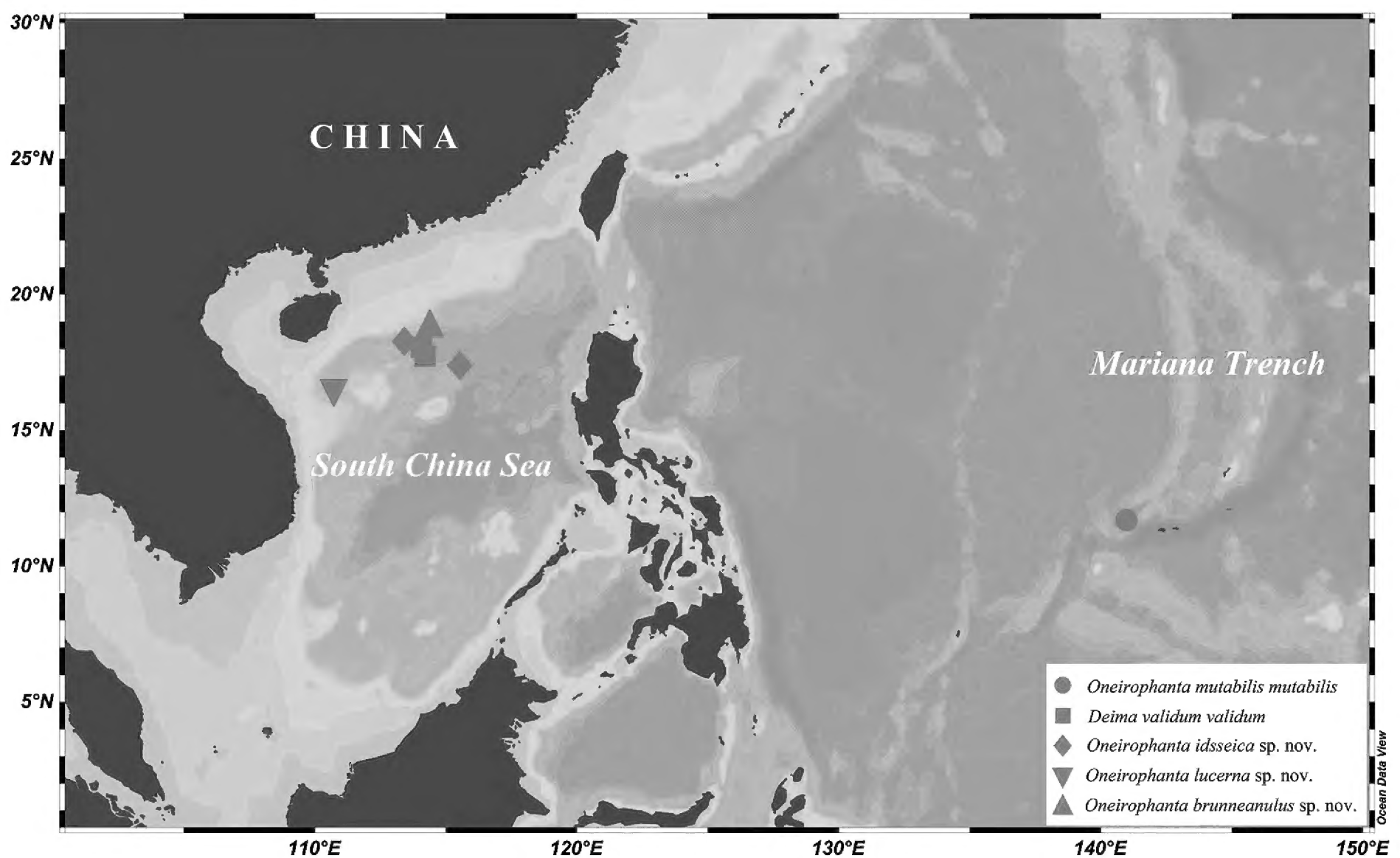


Figure 1. Sampling sites for examined species of Deimatidae in the South China Sea and the Mariana fore-arc area.

according to the manufacturer’s instructions. Mitochondrial cytochrome c oxidase I (COI) and 16S rRNA were generated for various specimens using the primers and methods outlined in Miller et al. (2017). The PCR amplification program was as follows: initial denaturation at 98 °C for 3 min, followed by 40 cycles at 98 °C for 10 s, 52 °C for 10s, 72 °C for 10s, and a final extension at 72 °C for 5 min. Total reaction volume was 50 µL: 25 µL Premix Taq with 1.25U Taq, 0.4 mM of each dNTP, 4 mMMg2+ (Ex Taq version, Takara, Dalian, China), 0.5 µM each of the primers and ~ 100 ng template DNA. The sequence chromatograms were then checked using CHROMAS 2.23 (Technelysium Pty Ltd.). The forward and reverse sequences were assembled using CONTIG EXPRESS, which is a component of Vector NTI Suite 6.0 (Life Technologies, Carlsbad, California).

Phylogenetic analyses

Two partial sequences (COI and 16S) were obtained from specimens and were deposited in GenBank (Table 1), and some relevant sequences (from all available deimatid species) that were downloaded from BOLD (<https://www.boldsystems.org/>) and NCBI (<https://www.ncbi.nlm.nih.gov/>) databases were used for phylogenetic analyses. *Apostichopus californicus* and *A. parvimensis* in the family Stichopodidae (order Synallactida) were used to root the tree.

Sequence alignments were generated using MAFFT7 (Kato and Standley 2013) with default parameters. Gblocks (Talavera and Castresana 2007) were used to remove batches of fragments from two alignments that were aligned ambiguously. The best partitioning scheme and evolutionary models for two pre-defined partitions were selected using PartitionFinder2 (Lanfear et al.

Table 1. List of GenBank accession numbers for all specimens used in this study.

Species	GenBank accession number		Reference
	16S	COI	
Deimatidae			
<i>Orphnurgus glaber</i>	KX856746	KX874361	Miller et al. 2017
<i>Deima validum</i>	KX856744	KX874364	Miller et al. 2017
<i>Deima validum validum</i> SY155-HS01	N/A	OR413734	this study
<i>Deima validum validum</i> SY84-HS02	OR658899	OR413743	this study
<i>Oneirophanta setigera</i>	KX856745	KX874363	Miller et al. 2017
<i>Oneirophanta</i> stet. CCZ_100	N/A	ON400706	Bribiesca-Contreras et al. 2022
<i>Oneirophanta</i> cf. <i>mutabilis</i>	ON406619	ON400724	Bribiesca-Contreras et al. 2022
<i>Oneirophanta idsseica</i> sp nov. SY86-HS01	OR658900	OR413744	this study
<i>Oneirophanta idsseica</i> sp. nov. SY84-HS01	OR658898	OR413742	this study
<i>Oneirophanta idsseica</i> sp. nov. SY283-HS01	OR658902	OR413737	this study
<i>Oneirophanta brunneannulata</i> sp. nov. SY157-HS01	OR658901	OR413733	this study
<i>Oneirophanta mutabilis</i> SY310-HS01	OR658897	OR413735	this study
<i>Oneirophanta lucerna</i> sp. nov. SY529-HS02	OR658906	OR413738	this study
<i>Oneirophanta lucerna</i> sp. nov. SY530-HS01	OR658903	OR413739	this study
<i>Oneirophanta lucerna</i> sp. nov. SY530-HS02	OR658904	OR413740	this study
<i>Oneirophanta lucerna</i> sp. nov. SY530-HS03	OR658905	OR413741	this study
Outgroups			
<i>Apostichopus californicus</i>	DQ777096	HM542319	Miller et al. 2017
<i>Apostichopus parvimensis</i>	KX856750	KX874373	Miller et al. 2017

2017), with all algorithms and AICc criteria. Maximum likelihood phylogenies (ML) were inferred using the Shimodaira-Hasegawa-like approximation likelihood-ratio test (Gascuel 2010) and IQ-TREE (Lam-Tung et al. 2015) models with 20,000 ultrafast bootstraps (Minh et al. 2013). Bayesian Inference phylogenies (BI) were inferred using MrBayes 3.2.6 (Ronquist et al. 2012) under the partition model (two parallel runs, 5,000,000 generations). The initial 25% of sampled data were discarded as burn-in, and the remaining trees were summarized in a 50% majority rule consensus tree. The results were visualized using FigTree v. 1.4.4. The Kimura two-parameter (K2P) genetic distances of COI among deimatid species were calculated using model MEGA X (Kumar et al. 2018).

Results

Taxonomy

Order Synallactida Miller, Kerr, Paulay, Reich, Wilson, Carvajal & Rouse, 2017

Family Deimatidae Théel, 1882

Genus Oneirophanta Théel, 1879

***Oneirophanta idsseica* sp. nov.**

<https://zoobank.org/50ADB642-A7E7-476A-AE6C-B45C2803F272>

Figs 2, 3

Oneirophanta stet. CCZ_100, Bribiesca-Contreras et al. 2022: 64–65, fig. 40.

Type material. Holotype. IDSSE-2018-0612-HS01, collected from the Xisha Trough of the South China Sea, station SY86-HS01 (18°16.11'N, 113°25.32'E), depth 2985 m, 12 Jun. 2018, preserved in absolute alcohol. **Paratypes.** Two specimens. IDSSE-2018-0531-HS01, collected from the Xisha Trough of the South China Sea, station SY84-HS01 (18°2.70'N, 114°3.51'E), depth 3156 m, 31 May 2018, preserved at -80 °C. IDSSE-2020-0917-HS01, collected from the northern slope of the South China Sea, station SY283-HS01 (17°23.20'N, 115°32.32'E), depth 3806 m, 17 Sep. 2020, preserved in absolute alcohol.

Type locality. In the Xisha Trough, which is located in the northern slope of the South China Sea, depth 2985 m.

Diagnosis. Body elongated and cylindrical, color yellowish-white. Tentacles 15. Ventrolateral tube feet up to 40–50 pairs, in alternating two or three rows. Dorsal papillae 18–20 on each side, in single rows. Ventrolateral papillae 9–12 on each side, in single rows. Midventral tube feet two and rudimentary. Dorsal deposits irregular perforated plates and varying types of crosses. Perforated plates and crosses with open ramifications ventrally. Papillae deposits slender and sturdy rods with spatulated ends, and crosses with open ramifications. Spatulated rods and irregular deposits in tube feet. Sturdy spatulated rods in tentacles.

Description of holotype. External morphology. Body elongated and cylindrical, ventrum flattened. 29 cm long and 9 cm wide before fixation (Fig. 2A, B). Color in vivo yellowish-white, tentacles, papillae, and tube feet often darker (Fig. 2C, D). Mouth anteroventral, anus posteroventral. Tentacle discs with rounded knobs at the edges, and the tentacles partially retracted into the mouth, making the number of tentacles difficult to calculate. Ventrolateral tube

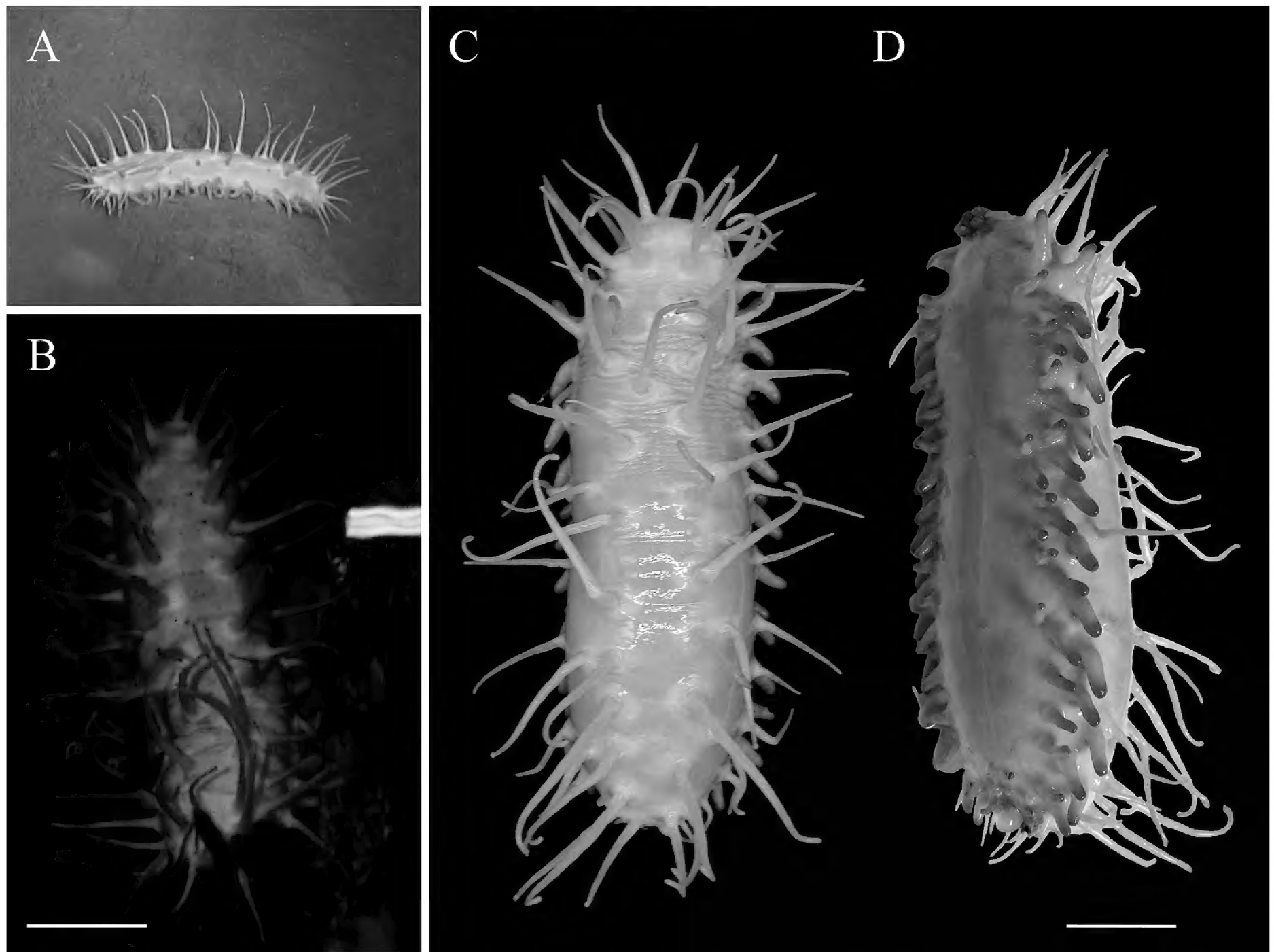


Figure 2. *Oneirophanta idsseica* sp. nov. (Holotype: IDSSE-2018-0612-HS01) **A** in situ image **B** holotype in live **C** dorsal view **D** ventrolateral view. Scale bars: 5 cm.

feet 40 pairs, arranged in alternating two or three rows on each ventrolateral ambulacrum. Midventral tube feet two and rudimentary, scattered along the mid-ventral ambulacrum, one positioned in the front third of the body, the other is positioned in the back third of the body, the anus surrounded by few small tube feet. Dorsal papillae roughly 18–20 on each body side, evenly distributed, measuring 4–10 cm in length, and placed in single rows along the dorsal radii. Ventrolateral papillae ~ 9–12 on each side, generally shorter than dorsal papillae, measuring 2.5–7 cm in length, arranged in single rows along the ventrolateral radii. All papillae slender and tapering from base to end.

Ossicle morphology. Dorsal deposits contain (1) perforated plates with open ramifications (Fig. 3A3, A9, A10), 0.3–0.8 mm in diameter, bearing 0–5 processes on the surface, central holes smaller towards the edge. (2) crosses two types, the first with dichotomously ramified ends, one side bearing 2–5 processes and the other side smooth and without spines (Fig. 3A5–A8); the second irregular, with arms slender than the first type and numerous bifurcated spines throughout the length (Fig. 3A1, A2). (3) spatulated rods with enlarged ends (Fig. 3A4). Papillae deposits contain (1) extremely slender or sturdy spatulated rods, 0.8–1.6 mm long (Fig. 3B1, B7–B10); (2) spatulated crosses with 2–3 processes on the surface (Fig. 3B2, B5, B6); (3) crosses with open ramifications resembling dorsal deposits (Fig. 3B3, B4). Deposits in tentacles only spatulated rods (Fig. 3C), more robust than spatulated rods in papillae. Tube

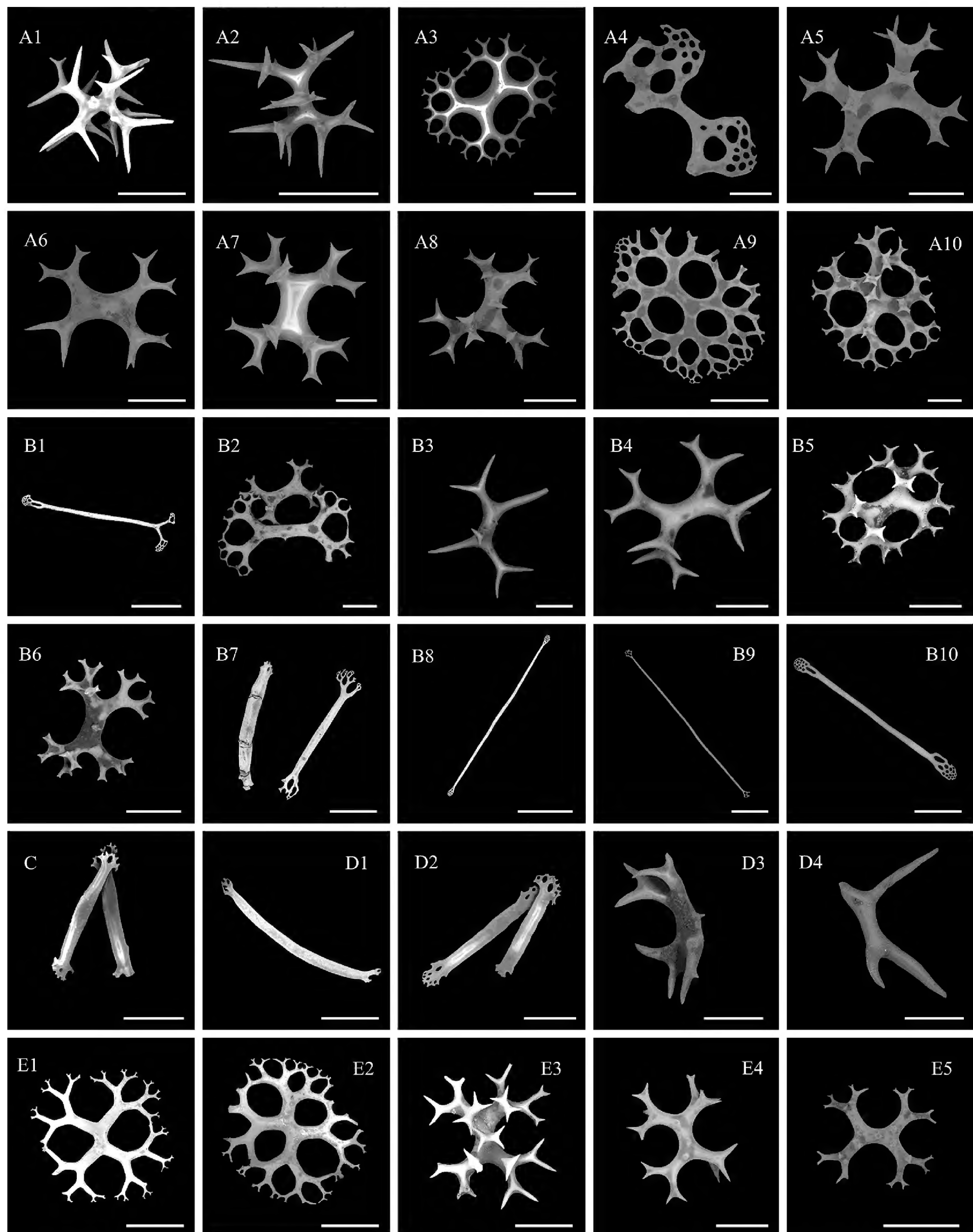


Figure 3. SEM images of different tissues from *Oneirophanta idsseica* sp. nov. (Holotype: IDSSE-2018-0612-HS01) **A1–A10** dorsal body wall **B1–B10** papillae **C** tentacles **D1–D4** tube feet **E1–E5** ventral body wall. Scale bars: 50 μ m (**A1, A2, A7, B3, B4, D3, D4, E4**); 100 μ m (**A3–A6, A8, A10, B2, B5, B6, E3, E5**); 300 μ m (**A9, B1, B7–B10, C, D1, D2, E1, E2**).

feet deposits mainly robust spatulated rods (Fig. 3D1, D2), 0.6–0.8 mm long, and irregular deposits, possibly in the developmental stage of perforated plates (Fig. 3D3, D4). Deposits in the ventrum same as dorsal (Fig. 3E1–E5), except for large spatulated rods with enlarged ends (Fig. 3A4).

Etymology. Consists of IDSSE and the Latin suffix *icus* (belonging to), to honor IDSSE's contributions and efforts to the field of deep-sea exploration.

Distribution. A seamount in APEI 4, Clarion Clipperton Zone; Xisha Trough, the northern slope of the South China Sea, at depths of 2985–3806 m.

Remarks. *Oneiophanta idsseica* sp. nov. is characterized by the arrangement of ventrolateral tube feet in two or three rows that number up to 40–50 pairs with distinctive cross-types in dorsal deposits. *O. idsseica* sp. nov. is distinct from *Oneiophanta setigera* (Ludwig 1893) due to the presence of small, perforated plates and crosses with open ramifications that are usually bifurcated. *O. idsseica* sp. nov. differs from *Oneiophanta conservata* Koehler & Vaney, 1905 and *Oneiophanta mutabilis* Théel, 1879 by the arrangement and high number of ventrolateral tube feet, and the absence of large, perforated plates on dorsum.

The phylogenetic trees showed that *O. idsseica* sp. nov., together with an unnamed species (*Oneiophanta* stet. CCZ_100, see below), formed a sister group that included *Oneiophanta* cf. *mutabilis* and *O. mutabilis*. From a morphological point of view, *O. idsseica* sp. nov. mostly resembled *O. stet.* CCZ_100 with ventrolateral tube feet arranged in two or three rows, two rudimentary midventral tube feet, spatulated crosses and small, irregular perforated plates on dorsum, and crosses with open ramifications in different stages of development on the ventrum. From a molecular point of view, the COI pairwise distance between *O. idsseica* sp. nov. and *O. stet.* CCZ_100 was 0.6% (Suppl. material 1). Compared with the description of *O. stet.* CCZ_100, our study provides more details of ossicle morphology of tentacles, tube feet, and papillae.

***Oneiophanta brunneannulata* sp. nov.**

<https://zoobank.org/97066926-74E7-4525-A25D-0F047F525BCC>

Figs 4, 5

Type material. Holotype. IDSSE-2018-0612-HS01, collected from the continental slope of the South China Sea, station SY157-HS01 (18°51.18'N, 114°24'E), depth 1340 m, 1 Jul. 2019, preserved in -80 °C

Type locality. On the continental slope of the South China Sea, depth 1340 m.

Diagnosis. Body elongated, color reddish brown, with darker tentacles and tube feet. Mouth and anus ventral. Tentacle 20. Ventrolateral tube feet ~ 37 pairs, each tube foot end with a brown ring, arranged in alternating three rows, bilateral symmetry. Dorsal papillae 23–26 on each body side, arranged in single rows. Ventrolateral papillae 9–11 on each body side. Midventral tube feet two and rudimentary. Deposits perforated plates, rods of varying shapes and few spatulated crosses.

Description of holotype. External morphology. Body elongated, dorsum convex, ventrum flattened. 20 cm long, and 5 cm wide before fixation (Fig. 4A, B). Skin reddish brown, with darker coloration on tentacles and tube feet, a brown ring at the end of each tube foot. Tentacle 20, never with ramified processes, circum-oral papillae absent. Ventrolateral tube feet ~ 37 pairs, placed in alternating three rows, bilateral symmetry. Midventral tube feet two and rudimentary, one placed on half the body, the other placed on a rear quarter of the body, and several small tube feet surrounding anus. Dorsal papillae 23–26 on each body side, arranged in single rows along dorsal radii, measuring 5–6 cm.

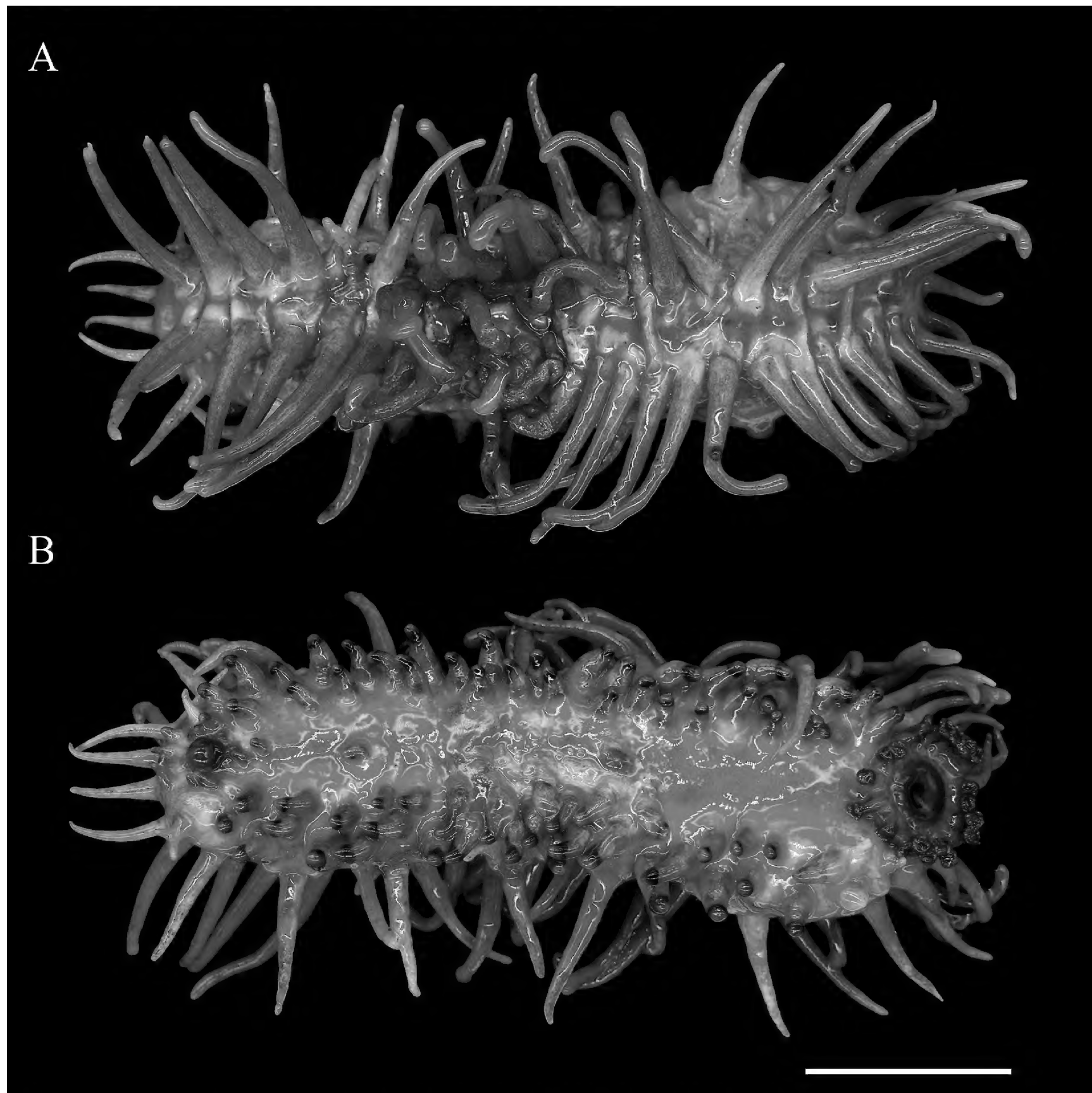


Figure 4. *Oneirophanta brunneannulata* sp. nov. (Holotype: IDSSE-2018-0612-HS01) **A** dorsal view **B** ventral view. Scale bar: 6 cm.

Ventrolateral papillae 9–11 on each side, arranged in single rows, comparatively shorter than dorsal papillae, measuring 2.3–4.6 cm.

Ossicle morphology. Dorsal deposits only robust perforated plates present (Fig. 5A1–A3), 0.6–1.1 mm in diameter, with central holes tapering from the center to the end, some perforated plates with irregular central apophysis (Fig. 5A1). Papillae contain (1) robust spatulated rods (Fig. 5B1–B3), 0.8–1 mm in length and partially connected at the ends (Fig. 5B1); (2) spatulated crosses (Fig. 5B4); (3) Perforated plates with 2–4 large central holes, rather slender than dorsal, measuring an average 0.9 mm in diameter (Fig. 5B5, B6). Tentacle deposits slender and sturdy rods with open ramifications (Fig. 5C), 0.4–0.6 mm in length. Tube feet with varying types of rods: (1) slender rods with open ramifications, 0.5–0.7 mm long (Fig. 5D1); (2) smooth spindle-shaped rods without spines, 0.7 mm in length (Fig. 5D2); (3) sturdy spatulated rods (Fig. 5D1, D3, D4), with rudimentary or enlarged ends, 0.5–0.9 mm in length. Numerous amorphous shaped, irregular broken deposits on ventrum (Fig. 5E1, E2).

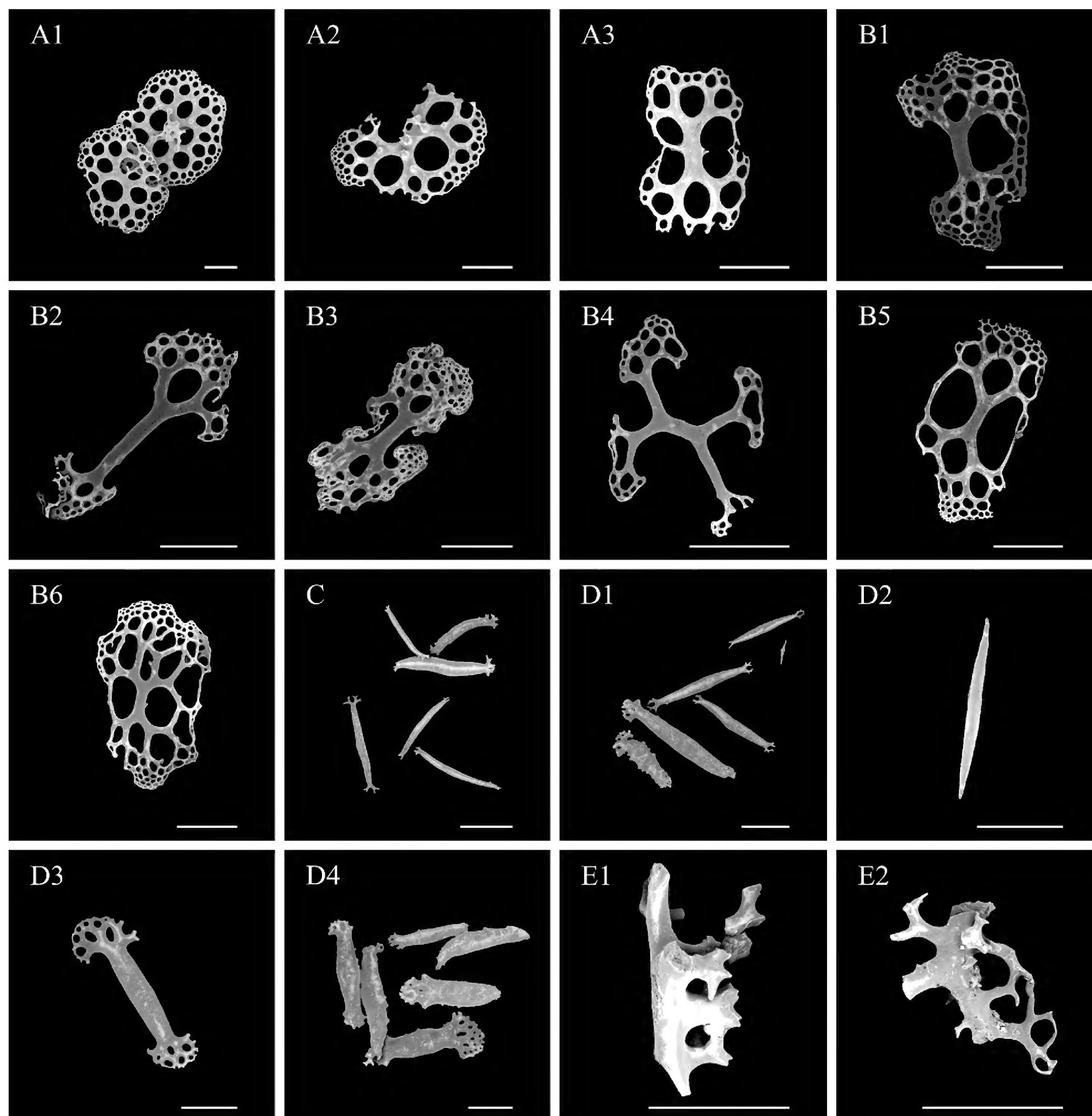


Figure 5. SEM images of *Oneiophanta brunneannulata* sp. nov. (Holotype: IDSSE-2018-0612-HS01) **A1–A3** dorsal body wall **B1–B6** papillae **C** tentacles **D1–D4** tube feet **E1–E2** ventral body wall. Scale bars: 300 μ m.

Etymology. The specific epithet *brunneannulata* in Latin means brown rings. It is here used as a noun in apposition and refers to the distinctive brown rings around the tube feet.

Distribution. Only in the type locality.

Remarks. *Oneiophanta brunneannulata* sp. nov. differs from other species in the genus in possessing brown rings at the end of tube feet that are arranged in three rows along ventrolateral radii. *O. brunneannulata* sp. nov. is relatively similar to *O. mutabilis* in possession of plates and spatulated rods, but there are differences: (1) different diameters of perforated plates, 0.6–1.1 mm in *O. brunneannulata* sp. nov., but 2–3 mm in *O. mutabilis*, and some perforated plates in *O. brunneannulata* sp. nov. possess a central apophysis. (2) different types of

deposits in tube feet, *O. brunneannulata* sp. nov. has various forms of spatulated rods and a few spindle-shaped rods, but lacks perforated plates, and there are perforated plates in addition to sturdy and spatulated rods in *O. mutabilis*. *O. brunneannulata* sp. nov. differs from *O. setigera* in having perforated plates on the dorsum and the ventrum, lacked spatulated crosses, and had a large number of tube feet that were arranged in three rows. Larger central perforations on perforated plates were in papillae, and the presence of spatulated rods and papillae were arranged in single rows along the dorsal radius (double rows along dorsal radius in *O. conservata*) distinguished *O. brunneannulata* sp. nov. from *O. conservata* (Table 2).

***Oneirophanta lucerna* sp. nov.**

<https://zoobank.org/DF0BA6C4-E07A-467A-9983-603B303CDA6E>

Figs 6, 7

Type material. Holotype. IDSSE-2023-0208-HS02, northern slope of the South China Sea, station SY530-HS02 (16°28.60'N, 110°18'E), depth 1389 m, 8 Feb. 2023, preserved in absolute alcohol. **Paratypes.** Three specimens. IDSSE-2023-0204-HS02, northern slope of the South China Sea, station SY529-HS02 (16°28.20'N, 110°43'E), depth 1393 m, 4 Feb. 2023, preserved in absolute alcohol. IDSSE-2023-0208-HS01, northern slope of the South China Sea, station SY530-HS01 (16°28.30'N, 110°43'E), depth 1389 m, 8 Feb. 2023, preserved in absolute alcohol. IDSSE-2023-0208-HS03, northern slope of the South China Sea, station SY530-HS03 (16°28.18'N, 110°43'E), depth 1392 m, 8 Feb. 2023, preserved in absolute alcohol.

Type locality. Northern slope of the South China Sea, depth 1389 m.

Diagnosis. Body elongated, skin smooth, color orange in vivo. Mouth anteroventral, anus posteroventral. Tentacle 19 or 20. Ventrolateral tube feet 11–14 on each body side, arranged in single rows. Midventral tube feet two and rudimentary. Dorsal papillae 15–27 on each side, placed in single rows along dorsal radius. Ventrolateral papillae 7–10 on each side, arranged in single rows. Dorsal deposits spatulated crosses, spatulated rods, and widely scattered spinous rods with branched spines. Papillae deposits with spinous rods and spatulated rods. Spatulated crosses with the arms twice divided and two types of spinous rods are in ventrum. Spinous rods and sturdy spatulated rods with open ramifications in tentacles and tube feet.

Description of holotype. External morphology. Body elongated, dorsum inflated, ventrum flattened, slightly narrowed anteriorly (Fig. 6). Length in vivo 22 cm (Fig. 6B), in ethanol 20 cm. Maximum body width 6 cm in vivo, 5.7 cm in ethanol. Color in vivo orange (Fig. 6C), tentacle and tips of papillae and tube feet more pigmented than the body skin. Tentacle 20. Circum-oral papillae absent. Mouth anteroventral, anus posteroventral terminal. Ventrolateral tube feet 14 pairs, placed in single rows on ventrolateral ambulacrum, partly projecting horizontally from the body (Fig. 6D). Midventral radius naked. Midventral tube feet two and rudimentary, with one placed on half the body, while the other placed on a rear quarter of the body, and several smaller tube feet close to anus. Left dorsal papillae 27, right dorsal papillae 21, measuring 1.5–5.2 cm, arranged in regular single rows along dorsal radii (Fig. 6C). Ventrolateral papillae 9–10 on each side, measuring 2.2–3.2 cm, placed in single rows along the ventrolateral radii.

Table 2. Main morphological characters in species of *Oneirophanta* Théel, 1879.

Characteristics	<i>O. idseica</i> sp. nov.	<i>O. brunneannulata</i> sp. nov.	<i>O. lucerna</i> sp. nov.	<i>O. conservata</i>	<i>O. setigera</i>	<i>O. mutabilis mutabilis</i>	<i>O. mutabilis affinis</i>
Tentacle number	15	20	19 or 20	Only eight founded	15–20	18–20	18–20
Ventrolateral tube feet number and arrangement	40–50 pairs, in alternating two or three rows	~ 37 pairs, in alternating three rows	11–14 on each side, in single rows	33–34 on each side, in two rows	16–30 on each side, in alternating double rows	8–28 (36), in alternating double rows	15–20 (44), –
Dorsal papillae number and arrangement	18–20 on each side, in single rows	23–26 on each body side, in single rows	15–27 on each body side, in regular single rows	30, in double rows	12–32, in irregular double rows	4–19, in single rows	5–35, in single or double rows
Ventrolateral papillae number and arrangement	9–12 on each side, in single rows	9–11, in single rows	7–10, in single rows	13, in two rows	9–17, –	4–17, –	5–11, –
Midventral tube feet number and arrangement	Two and rudimentary, one positioned in the front third of the body, the other positioned in the back third of the body	Two, rudimentary, one placed on half the body, the other placed on a rear quarter of the body	Two and rudimentary, one placed on half the body, the other on a rear quarter of the body	12, placed through-out the entire length of this radius, sometimes in pairs	0–6, in front of the anus	0–4, pre-anal	3–9, usually one pair placed pre-anal
Dorsal Deposit	Perforated plates with open ramifications and crosses with dichotomously ramified ends or irregular, and spatulated rods	Perforated plates, 0.6–1.1 mm in diameter, some with irregular central apophysis	Spatulated crosses, spinous rods with branched spines, spatulated rods up to 1 mm	Perforated plates, some in the developmental stage, rods with slightly thorny surfaces, pointed or bifid ends occasionally	Spatulated crosses, 1.1–3 mm	Perforated plates, 2–3 mm in diameter, bearing several small, vertical spines, with a rather slender mesh-work	Robust and rather small, vertical spines, often irregularly shaped due to elongation of the primary rod
Ventral	Perforated plates and crosses with open ramifications	Numerous irregular broken deposits, amorphous shaped	Spinous rods and spatulated crosses with arms twice divided	–	Spatulated crosses, 0.2–2.3 mm	Vary more, less well-developed, and less irregular than dorsal ones	–
Papillae	Slender or sturdy rods, crosses with open ramifications, some bearing 2–3 processes	Perforated plates with 2–4 large central holes, robust spatulated rods and few spatulated crosses	Spinous rods and spatulated rods		Spatulated rods	Perforated plates only	Perforated plates, small, sturdy and spatulated rods
Tube feet	Mainly robust spatulated rods and irregular deposits	Smooth spindle-shaped rods, slender rods with dichotomously ramified ends, sturdy spatulated rods with rudimentary or enlarged ends	Spinous rods of two types, one irregularly shaped, the other with few regularly distributed spines, sturdy spatulated rods with perforated extremities that occasionally bifurcated		–	Perforated plates	Sturdy and spatulated rods
Tentacle	Sturdy spatulated rods	Slender and sturdy rods with open ramifications			–	Irregularly placed and stout rods, somewhat branched	Clusters of rod-shaped spicules
Data source	Bribiesca-Contreras et al. 2022; this study	This study	This study	Koehler and Vaney 1905; Hansen 1975	Ludwig 1893; Hansen1975	Théel 1879; Hansen 1975	Théel 1879; Hansen 1975

‘–’ means data not available.

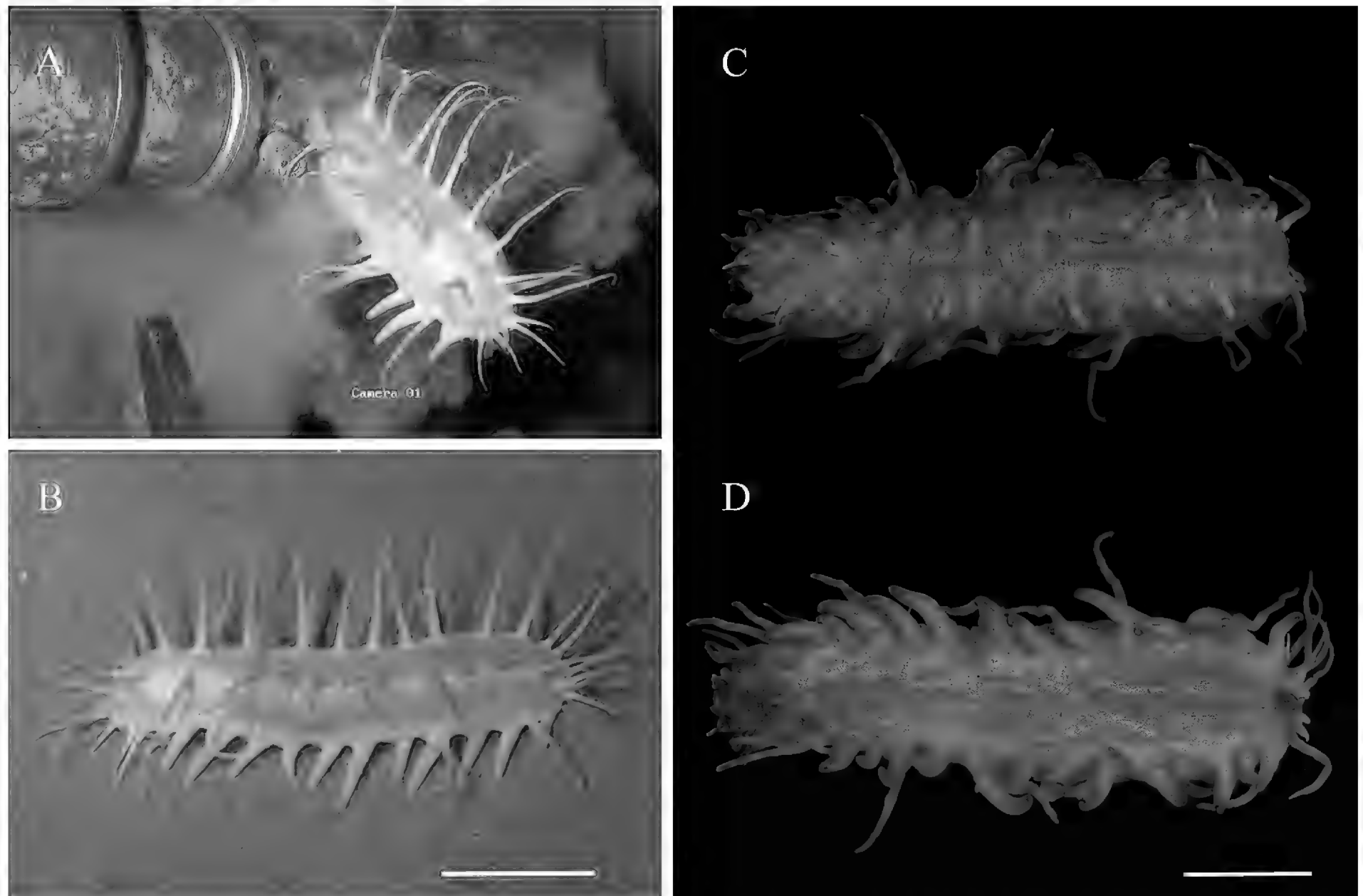


Figure 6. *Oneiophanta lucerna* sp. nov. (Holotype IDSSE-2023-0208-HS02) **A, B** in situ images **C** dorsal view **D** ventral view. Scale bars: 5 cm.

Ossicle morphology. Dorsal deposits contain spatulated crosses, spinous rods and spatulated rods. Spinous rods with irregular spines, 0.1–0.2 mm in length (Fig. 7A1, A2); spatulated crosses with arms 0.2–0.8 mm in length (Fig. 7A3); spatulated rods up to 1 mm (Fig. 7A4). Papillae with spinous rods 0.2–0.3 mm in length (Fig. 7B1, B2), and spatulated rods 0.4–0.9 mm in length, some with an extra branch from central part (Fig. 7B3), some bifurcated at the end (Fig. 7B4). Ventral body wall spatulated crosses with the arms twice divided (Fig. 7C1, C2) and the spinous rods of two types, one with more complex and irregularly placed spines (Fig. 7C3), the other with fewer and more regularly arranged spines (Fig. 7C4), up to 0.4 mm long. Tentacles and tube feet with similar ossicle types, spinous rods of two types, a few regularly distributed spines, 0.3–0.35 mm in length (Fig. 7D1, E1), irregularly shaped (Fig. 7D5, E2), and somewhat sturdy spatulated rods with perforated extremities that occasionally bifurcated (Fig. 7D2–D4, E3, E4), 0.2–0.7 mm in length.

Etymology. The species was named after the Latin word *lucerna* to commemorate the traditional Chinese Lantern Festival, which was relatively close to the time these specimens were collected.

Distribution. Northern slope of the South China Sea, depths of 1389–1393 m.

Remarks. The new species conformed to the genus *Oneiophanta* characterized by uncontracted the tentacles, the absence of oral papillae and tentacle discs without ramified processes. *Oneiophanta lucerna* sp. nov. differed from other species of *Oneiophanta* in possessing highly variable shaped spinous rods and ventrolateral tube feet that are only arranged in single rows, whereas,

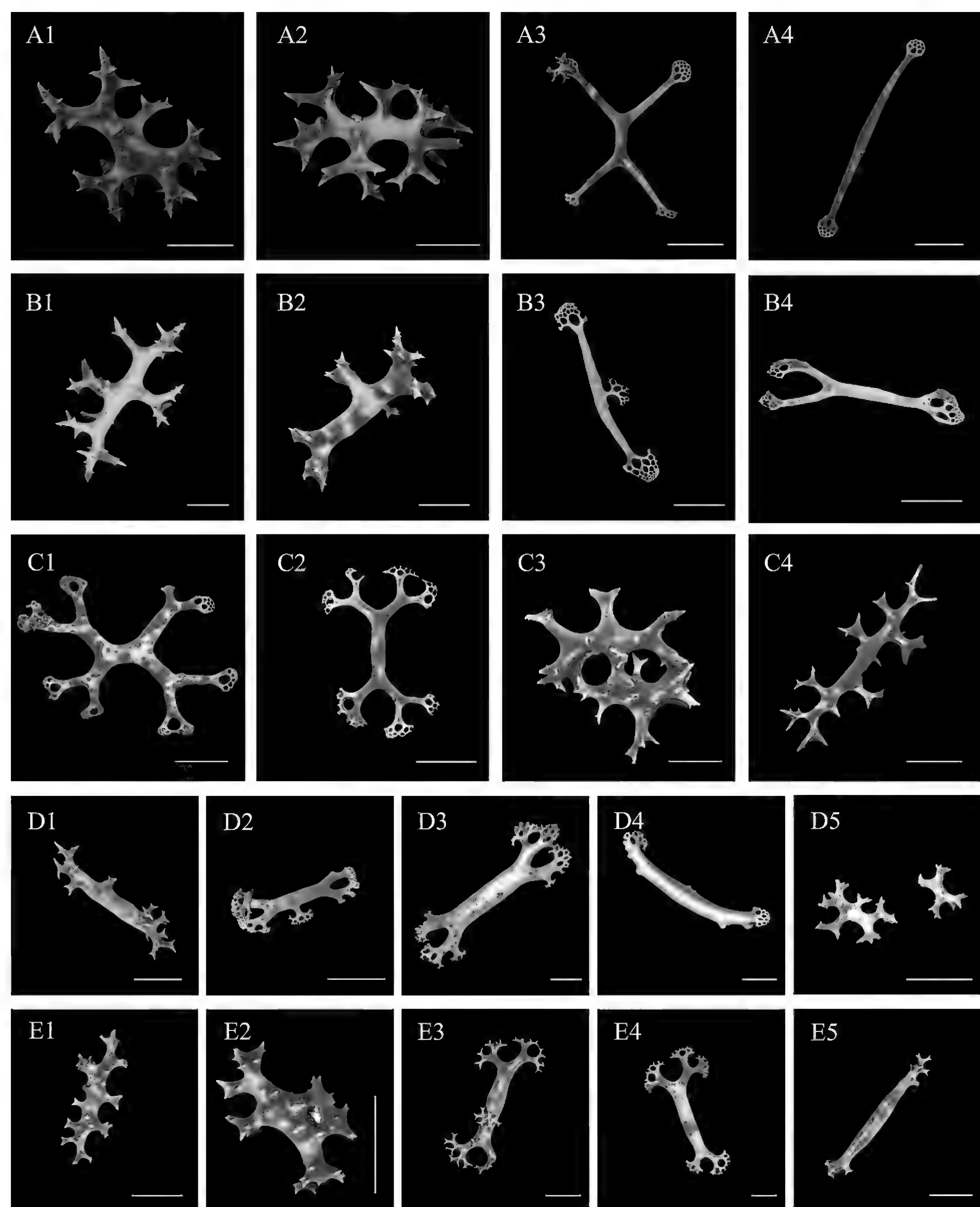


Figure 7. SEM images of *Oneirophanta lucerna* sp. nov. (Holotype IDSSE-2023-0208-HS02) **A1–A4** dorsal body wall **B1–B4** papillae **C1–C4** ventral body wall **D1–D5** tube feet **E1–E5** tentacles Scale bars: 50 μ m (**A1–A2**, **B1–B2**, **C2**); 100 μ m (**C3**, **D1–D5**); 200 μ m (**A3**, **A4**; **B3–C1**).

in other species (Table 2), they are arranged in two or three rows along ventro-lateral ambulacrum: *O. conservata*, *O. setigera*, and *O. mutabilis* in double rows, *O. idsseica* sp. nov. in alternating two or three rows, and *O. brunneannulata* sp. nov. in three rows. The three accepted species of *Oneirophanta* and the other

two new species in this study all have perforated plates that are absent in *Oneirophanta lucerna* sp. nov. This is the first record of a species in the genus *Oneirophanta* with mainly irregular spinous rods.

***Oneirophanta mutabilis mutabilis* Théel, 1879**

Figs 8, 9

Oneirophanta mutabilis Théel, 1879: 6–7, figs 4–6; Théel 1882: 62–68, pls XXI: 2, XXII, XXXI: 1–3, XXXVI: 1, 2, 8–11, XXXVII: 4, 13, XXXVIII: 11–12, XL: 1–3, XLI: 1, 2, 4, XLII: 9, XLIII: 1, 6, XLV, XLVI: 6, 7; R. Perrier 1902: 374–380, pl. XVIII: 10–15; Grieg 1921: pl. II: 1, 2; Hérouard 1923: 39–40, pls IV: 10, V: 3, 4; Agatep 1967: 63–65, pl. X: 1–7.

Oneirophanta mutabilis mutabilis Théel: Hansen 1967: 485–488, figs 3, 4; Hansen 1975: 24–32, figs 2–5.

Oneirophanta alternata R. Perrier, 1900: 117–118; R. Perrier 1902: 380–386, pls XIV: 3, 4, XVIII: 16–22.

Oneirophanta alternata var. *talismani* R. Perrier, 1902: 386–388, fig. 6.

Material examined. One specimen. IDSSE-2020-1203-HS01, in the Mariana fore-arc area, western Pacific Ocean, station SY310-HS01 (11°41.42'N, 140°58.56'E), depth 3394 m, 3 Dec. 2020, preserved in absolute alcohol.

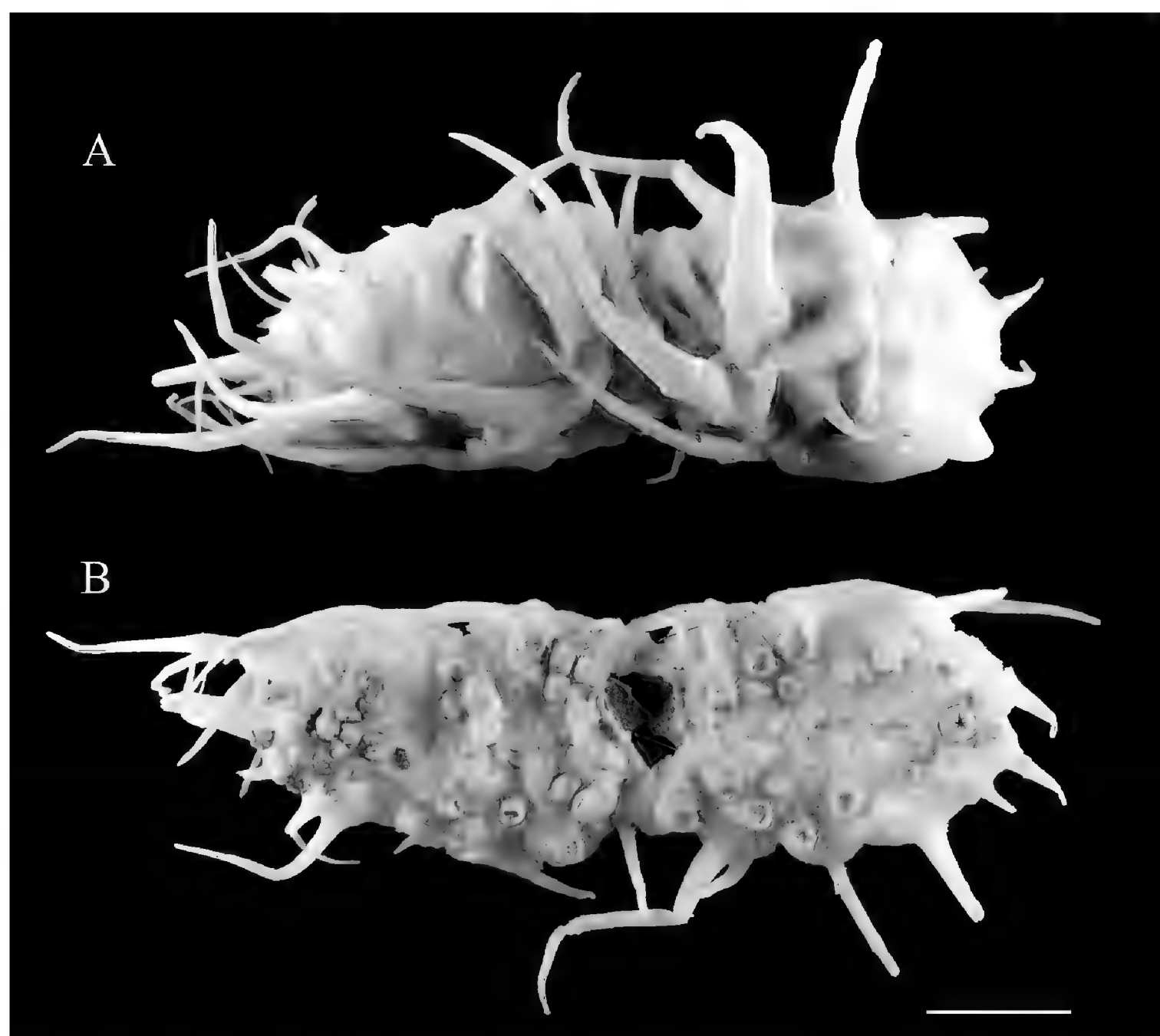


Figure 8. *Oneirophanta mutabilis mutabilis* Théel, 1879 (IDSSE-2020-1203-HS01) **A** dorsal view **B** ventral view. Scale bar: 3 cm.

Description. External morphology. Body cylindrical, nearly equal in width throughout the whole length and tapering anteriorly. 15 cm long and 4.5 cm wide after fixation with 95% alcohol for several days (Fig. 8A, B). Skin white, calcified and brittle. Tentacles 20, unretractile, 0.6–1.6 cm in length, with 7–9 marginal digits. Mouth and anus ventral. Ventrolateral ambulacra with 16 tube feet on each side, arranged in two irregular rows. Dorsal papillae 6 pairs, the maximum length up to 10 cm, placed in single rows along dorsal radii. Ventrolateral papillae 13 on each side, the maximum length up to 6 cm. Midventral tube feet not found due to damage to the mid-abdomen of the specimen, and only approximately six small tube feet observed near the anus.

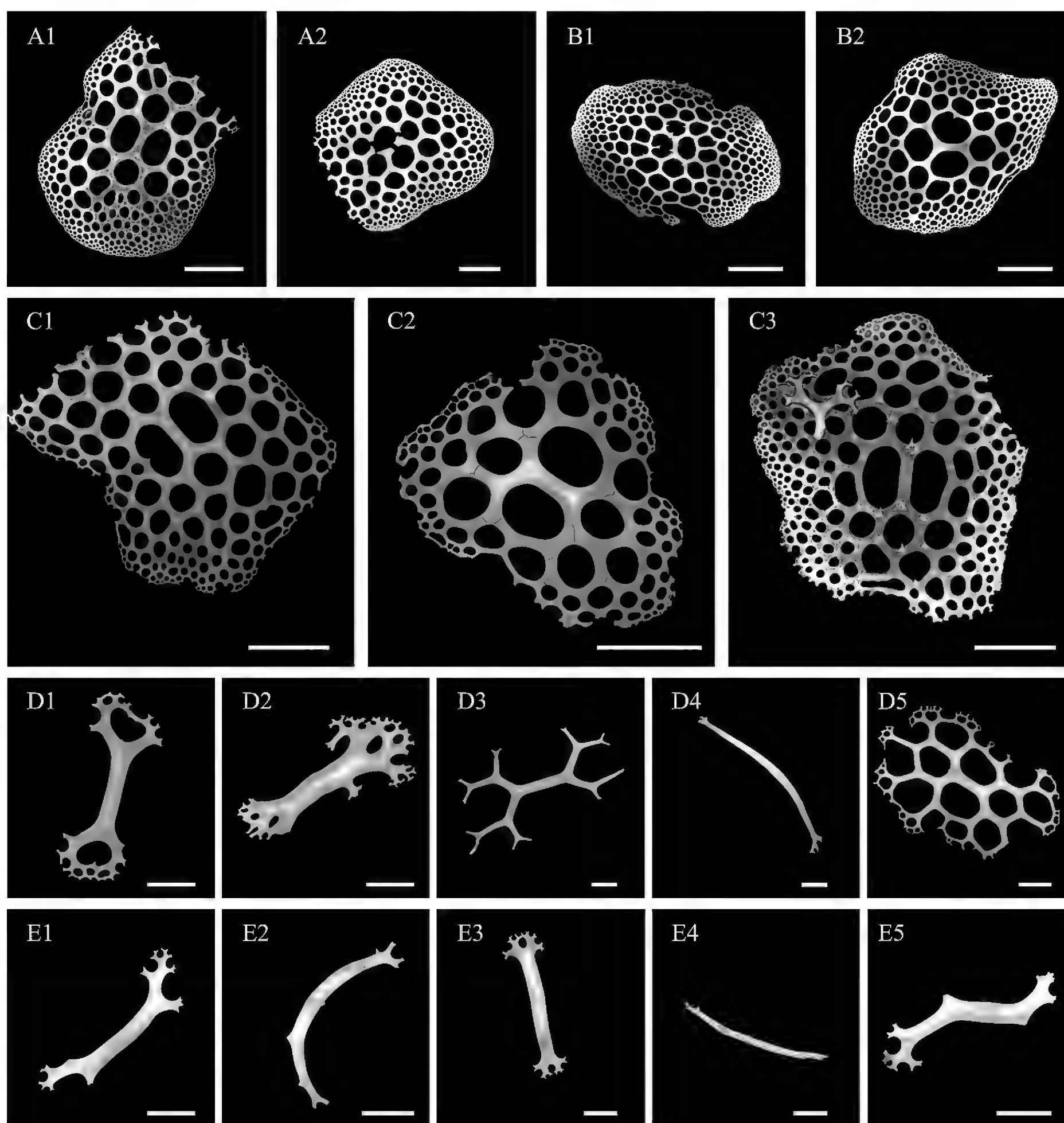


Figure 9. SEM images of *Oneirophanta mutabilis mutabilis* Théel, 1879 (IDSSE-20201203-HS01). **A1, A2** dorsal body wall **B1, B2** dorsal papillae **C1–C3** ventral body wall **D1–D5** tube feet **E1–E5** tentacle. Scale bars: 300 μ m (**A1–C3**); 100 μ m (**D1–E1, E3–E5**); 50 μ m (**E2**).

Ossicle morphology. The body wall and papillae with perforated plates (Fig. 9A1–C3), 0.9–1.5 mm in diameter. The entire periphery of fully developed plates usually surrounded by closed holes, with large central holes (especially in ventrum) and small peripheral holes. Some plates bearing 8–14 small, vertical spines (Fig. 9B1, C3), but no formation of a secondary layer of meshwork. Tube feet with rods up to 0.8 mm long (Fig. 9D1, D2, D4) and incompletely developed plates with open ramifications (Fig. 9D3, D5). The irregular rods in tentacles with variable bending angles (Fig. 9E1–E5).

Distribution. Cosmopolitan, depth 2515–6000 m (Hansen 1975, 1967; Thandar 1984).

Remarks. *Oneirophanta mutabilis* was first described west of the Crozet Islands (H.M.S. Challenger station 146: 46°46'S, 45°31'E) at depths of 2514 m (Théel 1879). It was divided into two subspecies, *Oneirophanta mutabilis mutabilis* Théel, 1879 and *Oneirophanta mutabilis affinis* Ludwig, 1893, based on differences in tentacle shape, the type of deposit, and the number of dorsal papillae (Hansen 1967). There are four main distinctions between them (see Table 2): (1) *O. m. affinis* was restricted to a relatively small area in the eastern Pacific where it replaced the otherwise cosmopolitan *O. m. mutabilis*. (2) tentacles with marginal digits were found in all specimens examined of *O. m. mutabilis*, but tentacle discs of *O. m. affinis* were smooth or had incised edges, and they lacked marginal digits. (3) the perforated plates of *O. m. affinis* were almost completely devoid of vertical spines, whereas those of *O. m. mutabilis* had vertical spines. (4) numerous rods of *O. m. affinis* were usually present in the tentacle discs, which was an additional difference from *O. m. mutabilis*. The morphological characteristics of our specimens were in accordance with the description of *O. m. mutabilis* (Hansen 1975). This was the first record of *O. m. mutabilis* from the Mariana fore-arc area.

Genus *Deima* Théel, 1879

Deima validum validum Théel, 1879

Figs 10, 11

Deima validum Théel, 1879: 5, figs 36–38; Théel 1882: 68–70, pls 18, 19, 31: 4–9, 36: 4, 37: 8, 43: 7, 44: 13, 46: 5; Sluiter 1901a: 60.

Deima validum validum: Hansen 1967: 488–490, fig. 5; Hansen 1975: 17–23, fig. 1, pls 11 (fig. 1), 13, (figs 1, 2); Bohn 2006: 9, fig. 4; Fernández-Rodríguez et al. 2019: 298, fig. 6.

Deima fastosum Théel, 1879: 5–6, figs 1–3; Théel 1882: 71–73, pls 20, 21: 1, 31: 10–13, 35: 7–10, 36: 7, 37: 3, 43: 2–3, 5, 46: 8.

Deima blakei Théel, 1886b: 1–2, figs 1, 2; Koehler and Vaney 1905: 55–57, pl. 11: 13–15; Hérouard 1923: 40–41, pls. 5: 7, 6: 5; Deichmann 1930: 115–116, pls 10: 7–11, 11: 1–3; Deichmann 1940: 198–199.

Deima atlanticum Hérouard, 1898: 88–89, figs 1, 2.

Deima mosaicum Ohshima, 1915: 233–234.

Material examined. Two specimens. IDSSE-2019-0630-HS01, collected from the northern slope of the South China Sea, station SY155-HS01 (17°43'N, 114°13'E), depth 3451 m, 30 Jun. 2019, preserved in absolute alcohol. IDSSE-2018-0531-

HS02, collected from the Xisha Trough of the South China Sea, station SY84-HS02 (18°2'N, 114°5'E), depth 3404 m, 31 May 2018, preserved at -80 °C.

Description. External morphology. Body ovate, dorsal vaulted, ventral flattened. 9–10 cm long and average 5.5 cm wide in vivo. Skin rigid, body wall brittle and easily broken. Color orange in vivo (Fig. 10A, B). Tentacles retracted into the mouth, resulting in uncountable quantities (Fig. 10C, D). 12 pairs of ventrolateral tube feet contractible based on morphology of different stages, single-rowed. Five pairs of rigid and conical dorsal papillae placed in two rows. Ventrolateral papillae three pairs, half the length of the body. Pre-anal tube feet absent.

Ossicle morphology. Basal layer and several additional layers amount in the center of the perforated plates on the body wall and dorsal papillae, in diameter 0.6–1.8 mm (Fig. 11A1–B3, D1, D2), with regular holes and a large, reticulated knob, ~ 0.2 mm high. The tube feet with some types of developmental stages towards perforated plates (Fig. 11C1, C3) and perforated plates with one layer (Fig. 11C2), rods bearing spines (Fig. 11C4).

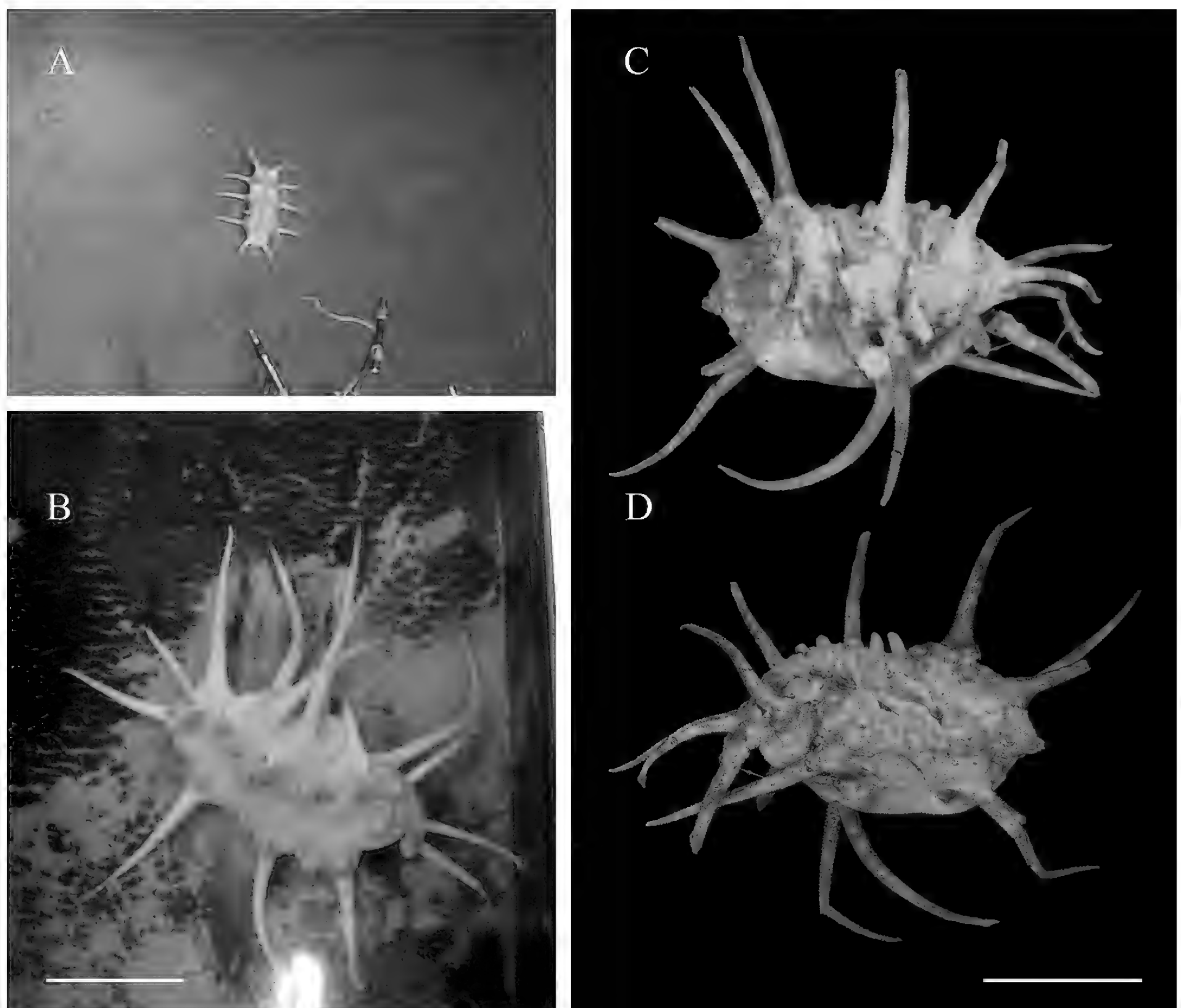


Figure 10. *Deima validum validum* (IDSSE-2018-0531-HS02 and IDSSE-2019-0630-HS01) **A** in situ image (IDSSE-2018-0531-HS02) **B** in vivo image (IDSSE-2018-0531-HS02) **C** dorsal view (IDSSE-2019-0630-HS01) **D** ventral view (IDSSE-2019-0630-HS01). Scale bars: 5 cm.

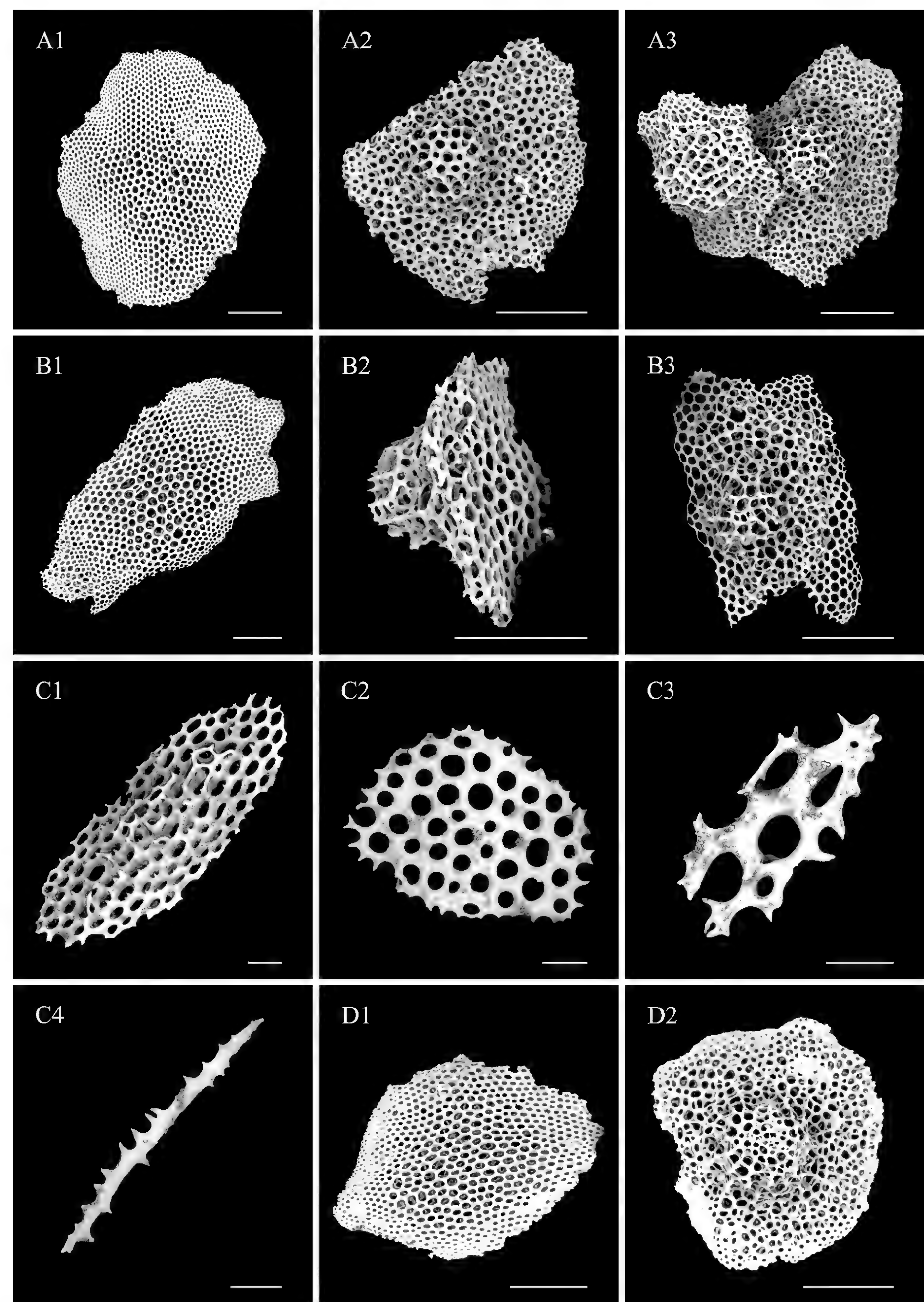


Figure 11. SEM images of *Deima validum validum* (IDSSE-2019-0630-HS01). **A1–A3** dorsal body wall **B1–B3** papillae **C1–C4** tube feet **D1–D2** ventral body wall. Scale bars: 300 μ m (**A1–B3**); 50 μ m (**D1, D2**)

Distribution. This subspecies probably has a cosmopolitan distribution, except for the Arctic and Southern Ocean, at depths of 724–5426 m (Théel 1882, 1886b; Sluiter 1901a; Hérouard 1902, 1923; Koehler and Vaney 1905; Ohshima 1915; Grieg 1921; Deichmann 1940; Hansen 1975; Sibuet 1977; O’Loughlin 1998; Bohn 2006).

Remarks. *Deima validum* was first described by Théel (1879) in the preliminary report of the exploring voyage of H. M. S. Challenger under Sir C. Wyville Thomson., with a detailed description of another novel species, *Deima fastosum*. Four new species were reported subsequently by different taxonomists: *D. atlanticum* Hérouard, 1898; *D. blakei* Théel, 1886; *D. mosaicum* Ohshima, 1915 and *D. pacificum* Ludwig, 1894. The family Deimatidae underwent a thorough revision by Hansen (1975), who regarded *D. fastosum*, *D. atlanticum*, *D. blakei*, *D. mosaicum* and *D. pacificum* as junior synonyms of *D. validum*. Hansen (1967) separated the species into two subspecies, cosmopolitan *Deima validum validum* and the eastern Pacific Ocean *Deima validum pacificum* Ludwig, 1894, based on differences in the number of dorsal papillae and the type of deposit in which they were found.

The two specimens examined here are consistent with the diagnosis of subspecies *D. v. validum* as described in detail by Hansen (1975). Perforated plates were like those of *D. fastosum*, with a large, reticulated, conical knob, which hardly ever approached in other known specimens that were used to investigate *D. v. validum*. This unique feature represents an extreme case of plate variation, where the spinous appearance of the skin is due to these very high and often vertically rising knobs on the plates. Hansen (1975) hypothesized that additional mesh structures on perforated plates varied with depth into two types: one-layered plates were characteristic of bathyal specimens, and many-layered plates were typical of abyssal specimens. However, it also exhibited a transitional type of plate in the abyssal Tasman Sea specimens and the bathyal Japanese specimens (perforated plates that were completely or almost completely devoid of additional meshwork), which prevented a clear distinction between a bathyal and an abyssal type of deposit.

Only in the Bay of Bengal (depth 1224–3365 m) did the development of additional layers of meshwork increase progressively with depth. In this research, high-knobbed plates were present in the abyssal South China Sea specimens (depth > 3000 m), but they were absent from the South China Sea specimens first reported (depth 1100 m) by Liao (1997). This was the second time that *Deima validum* has been recorded in the South China Sea. The proposition that the development of additional meshwork on perforated plates increased with depths needs to be investigated on more specimens at different depths in the South China Sea.

Genetic distance and phylogenetic analyses

The inter- and intraspecific genetic divergences of the COI gene were calculated to calculate the genetic distances in Deimatidae (Suppl. material 1). For the COI alignment, the interspecific distances in *Oneirophanta* ranged from 8.2–15.3%, the intraspecific distances in *Oneirophanta* were in the range of 0–0.6%, and the range of genetic distances among three genera was 15.1–21.9%.

In total, 11 COI sequences and 10 16S sequences were deposited into GenBank (Table 1). To check the intrageneric relationships of species in Deimatidae, a Bayesian phylogenetic tree and a Maximum likelihood (ML) tree were

reconstructed using concatenated 16S-COI sequences of length 1191 bp. The result of the phylogenetic analyses showed quite similar topologies in BI and ML trees (Fig. 12), except for one area of the BI tree, where *Oneirophanta setigera* formed an independent clade within *Oneirophanta* (BI 0.99).

The phylogenetic relationships of Deimatidae clustered into three portions and were consistent with the traditional classification system (Fig. 12). Portion 1: *Deima validum* was clustered with *Deima validum validum* (ML 99%, BI 1), which formed a monophyletic sister group (ML 89%, BI 1) with *Oneirophanta*. Portion 2: The three new species and the new record fell into *Oneirophanta*, which was divided into three clades. Clade 1: *O. setigera* was shown to be distinct from the other congeners in the BI tree (BI 0.99). But in the ML tree, *O. setigera* clustered with *O. brunneannulata* sp. nov., *O. cf. mutabilis*, *O. mutabilis* and *O. idsseica* sp. nov. with low support (ML 46%). Clade 2: *O. idsseica* sp. nov. clustered with *O. cf. mutabilis* and *O. mutabilis* (ML 49%, BI 0.79), followed by *O. brunneannulata* sp. nov. (ML 90%, BI 1). Clade 3: *O. lucerna* sp. nov. clustered with all other congeners in the ML tree (ML 69%). But in the BI tree, it was a sister taxon to species in clade 2, which then clustered with *O. setigera*. Portion 3: *Orphnurgus glaber* formed a separate clade with full node support (ML 100%, BI 1), and it was the only species in *Orphnurgus*.

Discussion

Species delineation and generic assignment

Both the morphology and molecular phylogenetic analyses supported the assignment of the three new species to the genus *Oneirophanta*. The external morphological characteristics in *Oneirophanta* species were quite similar to those in *Orphnurgus*, but *Oneirophanta* never has tentacle discs with ramified processes, and they usually have rounded knobs on the margin. The three new species described in this study conformed to this feature.

Oneirophanta brunneannulata sp. nov., *Oneirophanta idsseica* sp. nov. and *Oneirophanta lucerna* sp. nov. can be separated from other congeners by ossicle types, the arrangement and the number of dorsal papillae and tube feet. The separations were confirmed by the p-distance analyses, which showed that the uncorrected p-distance for the COI among *O. brunneannulata* sp. nov. and other congeners was 8.2–13.1%; among *O. idsseica* sp. nov. and other congeners was 8.2–14.1%, and among *O. lucerna* sp. nov. and other congeners was 12.1–14.8%. These divergences were much higher than the known intraspecific variation in *Oneirophanta* spp. (0–0.6%) (Suppl. material 1) and, thus, this warranted separation of *O. brunneannulata* sp. nov., *O. idsseica* sp. nov., and *O. lucerna* sp. nov. from other congeners.

The phylogenetic trees (Fig. 12) showed that *O. idsseica* sp. nov. clustered together with *Oneirophanta* stet. CCZ_100 from Clarion-Clipperton Zone, which was deposited in the Natural History Museum, London (voucher number: CCZ_100). Generally, taxonomic units with sequence differences of < 2% are likely to be the same species, and differences > 5% were confidently used to separate different species (Ward et al. 2008). Because the COI p-distance between the two species was 0.6% (Suppl. material 1), the divergences fell within the range of general intraspecific variation; both morphological characters and molecular data suggested that *O. idsseica* sp. nov. and *O. stet.* CCZ_100 are the same species.

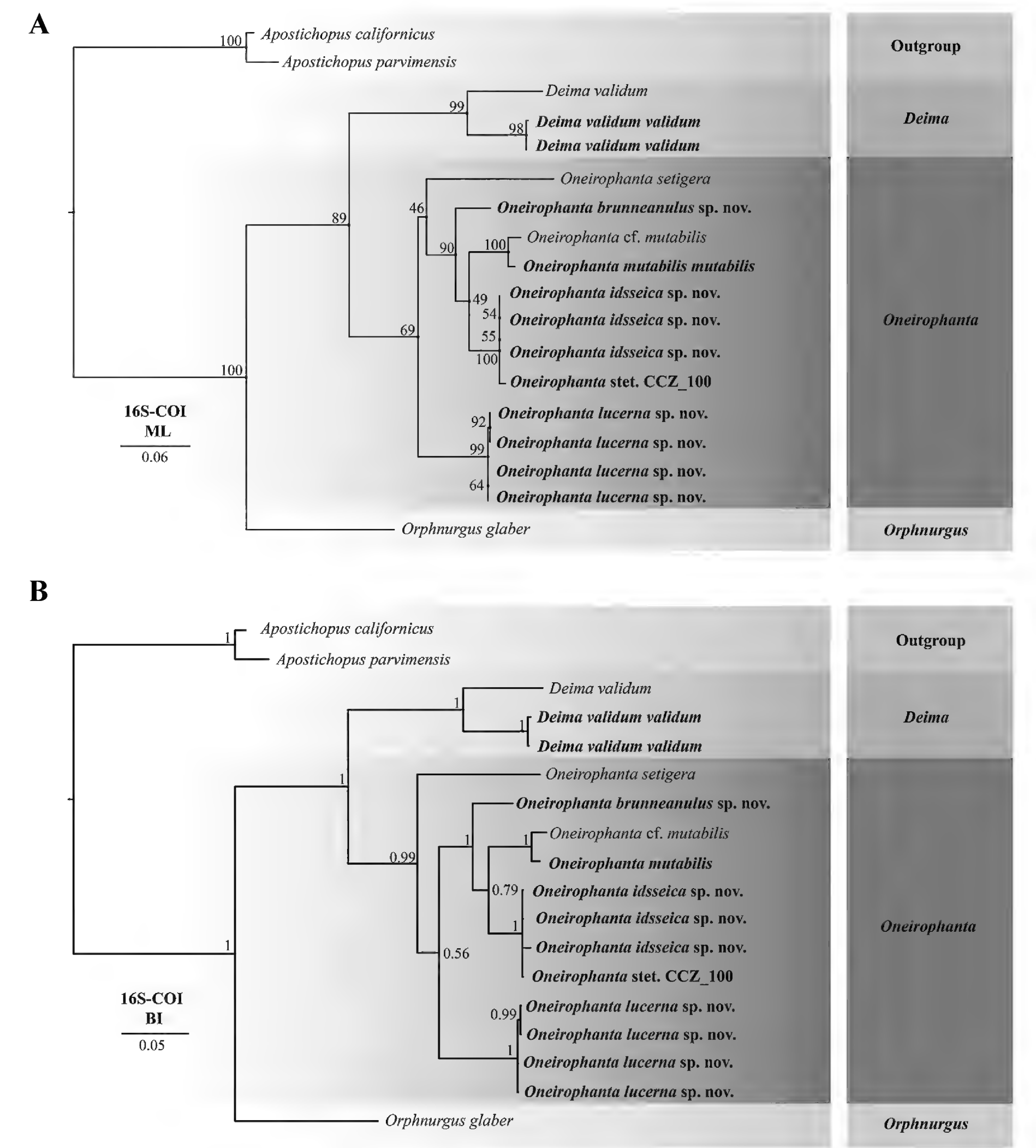


Figure 12. Maximum likelihood (ML) and Bayesian inference (BI) trees based on concatenated 16S-COI sequences showing phylogenetic relationships among deimatid species. The new sequences provided in this study are in bold **A** ML tree, with bootstrap replications labeled **B** BI tree, with posterior probability labeled.

Geographic distribution of deimatid species

There are a total of three genera and 16 species in the family Deimatidae, which include the three species that we described here. To date, 11 species of Deimatidae have been discovered in the deep water of the Pacific Ocean. *Deima* only includes one species: *Deima validum*, which occurs worldwide at depths of 724–5426 m. Nine species of *Orphnurgus* are accepted, with five species recorded from the Pacific: *Orphnurgus dorisae* Pawson, 2002 from the southern Pacific

Ocean (New Zealand), *Orphnurgus glaber* Walsh, 1891 from the central and western Pacific Ocean, *Orphnurgus protectus* (Sluiter 1901) and *Orphnurgus bacillus* Cherbonnier & Féral, 1981 from the western Pacific Ocean (Celebes Strait and Philippines), and *Orphnurgus vitreus* (Fisher 1907) from the North Pacific Ocean (off Hawaiian Islands). On a vertical scale, all species in this genus inhabited the water from relatively shallow depths to the bathyal zone (depth 174–1301 m). Among the six species, which included the three new species of *Oneirophanta*, *Oneirophanta mutabilis*, the type species of the genus, is a cosmopolitan species (Hansen 1975). *O. conservata* was the only species recorded from the Indian Ocean (Koehler and Vaney 1905). All the other species were found mainly in the Pacific Ocean: *O. setigera* from the southern and eastern Pacific (Kermadec Trench and Gulf of Panama), *O. mutabilis mutabilis* is a cosmopolitan species with multiple records in the Pacific Ocean (Hansen 1975), and *O. mutabilis affinis* is restricted to a relatively small area in the eastern Pacific Ocean. The three new species were from the western Pacific Ocean (South China Sea). In addition, *O. conservata* has the shallowest record of the genus (depth 1315 m), and *O. mutabilis* has the deepest record (depth 6000 m). *Oneirophanta* was distinguished from the other two genera by its inhabitation in the bathyal-abyssal zone.

Based on their distribution, deimatid holothurians are abundant in the Pacific Ocean and inhabit a wide range of depths (174–6000 m). Future expeditions to the Pacific zone may discover even more species, and more research is needed to evaluate the species diversity and geographic distribution of these deep-sea holothurians.

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

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Author contributions

Yunlu Xiao conceived and designed this project, performed morphological examination and description, conducted molecular analyses, and wrote or reviewed drafts of the paper. Haibin Zhang conceived and designed this project, reviewed and edited drafts of the paper and approved the final draft.

Author ORCIDs

Haibin Zhang  <https://orcid.org/0000-0001-5429-9851>

Data availability

All of the data that support the findings of this study are available in the main text or Supplementary Information.

References

- Agatep CP (1967) Some elasipodid holothurians of Antarctic and Subantarctic seas. Antarctic Research Series III 11: 49–71. <https://doi.org/10.1029/AR011p0049>
- Billett DM (1991) Deep Sea Holothurians. Oceanography and Marine Biology 29: 259–317.
- Bohn J (2006) Crinoidea and Holothuroidea (Echinodermata) of the abyssal Angola Basin Results of the DIVA-1 expedition of FS “Meteor” (Cruise M48/1). Zootaxa 1276(1): 1–31. <https://doi.org/10.11646/zootaxa.1276.1.1>
- Bribiesca-Contreras G, Dahlgren TG, Amon DJ, Cairns S, Drennan R, Durden JM, Eléaume MP, Hosie AM, Kremenetskaia A, McQuaid K, O’Hara TD, Rabone M, Simon-Lledó E, Smith CR, Watling L, Wiklund H, Glover AG (2022) Benthic megafauna of the western Clarion-Clipperton Zone, Pacific Ocean. ZooKeys 2022: 1–110. <https://doi.org/10.3897/zookeys.1113.82172>
- Cherbonnier G, Féral JP (1981) Echinodermes: Holothuries. In: Forest J (Ed.) Résultats des campagnes MUSORSTOM: 1. Philippines (18–28 Mars 1976). Mémoires du Muséum national d’Histoire naturelle. Série A, Zoologie 91: 357–412.
- Deichmann E (1930) The holothurians of the western part of the Atlantic Ocean. Bulletin of the Museum of Comparative Zoology at Harvard College 71(3): 43–226.
- Deichmann E (1940) Report on the Holothurians, collected by the Harvard-Havana Expeditions 1938 and 1939, with a revision of the Molpadonia of the Atlantic Ocean. Memorias de la Sociedad Cubana de Historia Natural ‘Felipe Poey’ 14: 183–240.
- Fernández-Rodríguez I, Arias A, Anadón N, Acuña JL (2019) Holothurian (Echinodermata) diversity and distribution in the central Cantabrian Sea and the Avilés Canyon System (Bay of Biscay). Zootaxa 4567(2): 293–325. <https://doi.org/10.11646/zootaxa.4567.2.5>
- Fisher WK (1907) The Holothurians of the Hawaiian Islands. Proceedings of the United States National Museum 32(1555): 637–744. <https://doi.org/10.5479/si.00963801.32-1555.637>
- Gascuel O (2010) New Algorithms and Methods to Estimate Maximum-Likelihood Phylogenies: Assessing the Performance of PhyML 3.0. Systematic Biology 59(3): 307–321. <https://doi.org/10.1093/sysbio/syq010>
- Grieg JA (1921) Echinodermata. Report on the scientific results of the “Michael Sars” North Atlantic deep-sea expedition 1910 3(2): 1–47. <https://doi.org/10.5962/bhl.title.79346>
- Haeckel E (1896) Systematische Phylogenie der Echinodermen. Systematische Phylogenie der Wirbellosen Tiere (Invertebrata): Zweiter Teil des Entwurfs einer systematischen Stammengeschichte. Reimer, Berlin, 348–504. <https://doi.org/10.1515/9783111443935>

- Hansen B (1967) The taxonomy and zoogeography of the deep-sea holothurians, in their evolutionary aspects. *Studies in Tropical Oceanography* 5: 480–501.
- Hansen B (1975) Scientific results of the Danish Deep-Sea Expedition Round the World 1950–52. Systematics and biology of the deep-sea holothurians. Part 1: Elasipoda. *Galathea Report* 13: 1–262.
- Hendler G, Miller JE, Pawson DL, Kier PM (1995) Sea stars, sea urchins, and allies: echinoderms of Florida and the Caribbean.
- Hérourard E (1898) Note préliminaire sur les Holothuries provenant des dragages du yacht "Princesse-Alice". *Bulletin de la Société Zoologique de France* 23: 88–89.
- Hérourard E (1902) Holothuries provenant des campagnes de la Princesse-Alice (1892–1897). Résultats des campagnes scientifiques accomplies sur son yacht par Albert Ier. *Prince Souverain de Monaco* 21: 1–61.
- Hérourard E (1923) Holothuries provenant des campagnes des yachts PRINCESSE-ALICE et HIRONDELLE II (1898–1915). Résultats des campagnes scientifiques accomplies sur son yacht par Albert Ier. *Prince Souverain de Monaco* 66: 1–163.
- Katoh K, Standley D (2013) MAFFT Multiple Sequence Alignment Software Version 7: Improvements in performance and usability. *Molecular Biology and Evolution* 30(4): 772–780. <https://doi.org/10.1093/molbev/mst010>
- Koehler R, Vaney C (1905) An Account of the Deep-Sea Holothuroidea Collected by the Royal Indian Marine Survey Ship Investigator. The Indian Museum, Calcutta, 1–170. <https://doi.org/10.5962/bhl.title.1730>
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K (2018) MEGA X: Molecular evolutionary genetics analysis across computing platforms. *Molecular Biology and Evolution* 35(6): 1547–1549. <https://doi.org/10.1093/molbev/msy096>
- Lam-Tung N, Schmidt HA, Arndt VH, Quang MB (2015) IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molecular Biology and Evolution* 32(1): 268–274. <https://doi.org/10.1093/molbev/msu300>
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B (2017) PartitionFinder 2: New methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. *Molecular Biology and Evolution* 34: 772–773. <https://doi.org/10.1093/molbev/msw260>
- Li XZ (2017) Taxonomic research on deep-sea macrofauna in the South China Sea using the Chinese deep-sea submersible Jiaolong. *Integrative Zoology* 12(4): 270–282. <https://doi.org/10.1111/1749-4877.12254>
- Li YN, Xiao N, Zhang LP, Zhang H (2018) *Benthodytes marianensis*, a new species of abyssal elasipodid sea cucumbers (Elasipodida: Psychropotidae) from the Mariana Trench area. *Zootaxa* 4462(3): 443–450. <https://doi.org/10.11646/zootaxa.4462.3.10>
- Li XZ, Dong D, Kou Q, Yang M, Gong L, Sui JX (2019) Advances in research on deep-sea macrobenthic biodiversity with the progress in China. *Acta Oceanologica Sinica* 41(10): 169–181.
- Liao YL (1997) Fauna Sinica. Phylum Echinodermata. Class Holothuroidea. Science Press, Beijing, 334 pp.
- Liao YL, Xiao N (2012) Species composition and faunal characteristics of echinoderms in China seas. *Shengwu Duoyangxing* 19(6): 729–736. <https://doi.org/10.3724/SP.J.1003.2011.08155>
- Ludwig H (1893) Vorläufiger bericht über die erbeuteten holothurien. Reports on the Dredging Operations off the West Coast of Central America to the Galapagos, etc., by the U.S. Fish Commission Steamer "Albatross". IV. *Bulletin of the Museum of Comparative Zoölogy at Harvard College* 24(4): 105–114.

- Ludwig H (1894) The Holothurioidea. Reports on an exploration off the west coasts of Mexico, Central and South America, and off the Galapagos Islands, in charge of Alexander Agassiz, by the U.S. Fish Commission Steamer "Albatross", during 1891. *Memoirs of the Museum of Comparative Zoology at Harvard College* 17(3): 1–183.
- Miller AK, Kerr AM, Paulay G, Reich M, Wilson NG, Carvajal JI, Rouse GW (2017) Molecular phylogeny of extant Holothuroidea (Echinodermata). *Molecular Phylogenetics and Evolution* 111: 110–131. <https://doi.org/10.1016/j.ympev.2017.02.014>
- Minh BQ, Nguyen MAT, Von Haeseler A (2013) Ultrafast Approximation for Phylogenetic Bootstrap. *Molecular Biology and Evolution* 30(5): 1188–1195. <https://doi.org/10.1093/molbev/mst024>
- Morton B, Blackmore G (2001) South China Sea. *Marine Pollution Bulletin* 42(12): 1236–1263. [https://doi.org/10.1016/S0025-326X\(01\)00240-5](https://doi.org/10.1016/S0025-326X(01)00240-5)
- O'Loughlin PM (1998) Elapod holothurians from the continental slope of Australia. *Echinoderms*: 499–504.
- Ohshima H (1915) Report on the holothurians collected by the United States Fisheries Steamer "Albatross" in the northwestern Pacific during the summer of 1906. *Proceedings of the United States National Museum* 48(2073): 213–291. <https://doi.org/10.5479/si.00963801.48-2073.213>
- Pawson DL (1982a) Holothuroidea. Synopsis and classification of living organisms, vol. 2. McGraw-Hill, New York, 1232 pp.
- Pawson DL (2002) A new species of bathyal elapod sea cucumber from New Zealand (Echinodermata: Holothuroidea). *New Zealand Journal of Marine and Freshwater Research* 36(2): 333–338. <https://doi.org/10.1080/00288330.2002.9517090>
- Perrier R (1900) Diagnose des espèces nouvelles d'Holothuries draguées par le Travailleur et le Talisman. (Troisième et dernière Note.). *Bulletin du Muséum National d'Histoire Naturelle* 6: 116–119. <https://doi.org/10.5962/bhl.part.12287>
- Perrier R (1902) Holothuries. *Expéditions scientifiques du Travailleur et du Talisman pendant les années 1880, 1881, 1882, 1883*, Paris, 273–554.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MAHJ, Huelsenbeck JP (2012) MrBayes 3.2. *Systematic Biology* 61(3): 539–542. <https://doi.org/10.1093/sysbio/sys029>
- Sibuet M (1977) Distribution and diversity of echinoderms (Holothuroidea, Asteroidea) in the abyssal zone in the Bay of Biscay. *Deep-Sea Research* 24: 549–564. [https://doi.org/10.1016/0146-6291\(77\)90527-6](https://doi.org/10.1016/0146-6291(77)90527-6)
- Sluiter CP (1901a) Die Holothurien der Siboga-Expedition. *Biodiversity Heritage Library OAI Repository* 44: 1–142. <https://doi.org/10.5962/bhl.title.85348>
- Sluiter CP (1901b) Neue holothurien aus der Tiefsee des Indischen Archipels Gesammelt durch die Siboga-Expedition. *Tijdschrift der Nederlandsche Dierkundige Vereeniging* 7(1): 1–28.
- Smirnov A (2012) System of the class Holothuroidea. *Paleontological Journal* 46(8): 793–832. <https://doi.org/10.1134/S0031030112080126>
- Talavera G, Castresana J (2007) Improvement of Phylogenies after Removing Divergent and Ambiguously Aligned Blocks from Protein Sequence Alignments. *Systematic Biology* 56(4): 564–577. <https://doi.org/10.1080/10635150701472164>
- Teh LSL, Cashion T, Alava Saltos JJ, Cheung WWL, Sumaila UR (2019) Status, Trends, and the Future of Fisheries in the East and South China Seas. *Fisheries Centre Research Reports* 27(1): 1–101.
- Thandar AS (1984) The holothurian fauna of southern. PhD Thesis. University of Durban-Westville, 566 pp.

- Théel H (1879) Preliminary Report on the Holothuridae of the Exploring Voyage of H.M.S. “Challenger” under Professor Sir C. Wyville Thomson F.R.S., Part 1, Bihang Till K. Svenska Vetenskaps-Akademiens Handlingar 5(19): 1–20.
- Théel H (1882) Report on the Holothuroidea. Part I. Report of the scientific results of the voyage of H.M.S. Challenger. Zoology : Analysis of Complex Systems, ZACS 4(13): 1–176.
- Théel H (1886b) Report on the Holothuroidea. Reports on the results of dredging. “Blake”, 30. Bulletin of the Museum of Comparative Zoology at Harvard 13(1): 1–21.
- Walsh JHT (1891) List of deep-sea holothurians collected during seasons 1887 to 1891, with description of new species. Natural History Notes from HM. Indian Survey Steamer INVESTIGATOR, Commander RF. Hoskyn, RN. commanding. No. 24. Journal of the Asiatic Society of Bengal 60: 197–204.
- Ward RD, Holmes BH, O’Hara TD (2008) DNA barcoding discriminates echinoderm species. Molecular Ecology Resources 8(6): 1202–1211. <https://doi.org/10.1111/j.1755-0998.2008.02332.x>
- WoRMS (2023) Deimatidae Théel, 1882. <https://www.marinespecies.org/aphia.php?p=taxdetails&id=123190> [Accessed on 19 Oct 2023]
- Xiao N, Li XM, Sha ZL (2018) Psychropotid holothurians (Echinodermata: Holothuroidea: Elasipodida) of the tropical Western Pacific collected by the KEXUE expedition with description of one new species. Marine Biology Research 14(8): 816–826. <https://doi.org/10.1080/17451000.2018.1546012>
- Xiao YL, Xiao N, Zeng XQ (2023) *Benthodytes tetrapapillata* sp. nov., a new elasipodid sea cucumber (Elasipodida: Psychropotidae) from a seamount in the Western Pacific Ocean. Journal of Oceanology and Limnology 41(5): 1978–1987. <https://doi.org/10.1007/s00343-022-2220-0>

Supplementary material 1

Estimates of p-distances of the COI gene among deimatid species and studied sequences

Authors: Yunlu Xiao, Haibin Zhang

Data type: docx

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